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Hybrid nanostructures for neurodegenerative disease theranostics: the art in the combination of biomembrane and non-biomembrane nanostructures

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Abstract

The diagnosis of neurodegenerative diseases (NDDs) remains challenging, and existing therapeutic approaches demonstrate little efficacy. NDD drug delivery can be achieved through the utilization of nanostructures, hence enabling multimodal NDD theranostics. Nevertheless, both biomembrane and non-biomembrane nanostructures possess intrinsic shortcomings that must be addressed by hybridization to create novel nanostructures with versatile applications in NDD theranostics. Hybrid nanostructures display improved biocompatibility, inherent targeting capabilities, intelligent responsiveness, and controlled drug release. This paper provides a concise overview of the latest developments in hybrid nanostructures for NDD theranostics and emphasizes various engineering methodologies for the integration of diverse nanostructures, including liposomes, exosomes, cell membranes, and non-biomembrane nanostructures such as polymers, metals, and hydrogels. The use of a combination technique can signifcantly augment the precision, intelligence, and efficacy of hybrid nanostructures, therefore functioning as a more robust theranostic approach for NDDs. This paper also addresses the issues that arise in the therapeutic translation of hybrid nanostructures and explores potential future prospects in this feld.

Keywords Neurodegenerative diseases, Biomembrane, Hybrid nanostructure, Alzheimer's disease, Parkinson's disease, Diagnosis

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Introduction

Neurodegenerative diseases (NDDs) are a diverse group of neurological disorders characterized by the progressive loss of neurons in the peripheral nervous system or the central nervous system (CNS), leading to cognitive and behavioral deficits $[1]$ $[1]$. These diseases include amyotrophic lateral sclerosis, Alzheimer's disease (AD), multiple sclerosis (MS), Parkinson's disease (PD), and Huntington's disease (HD). The prevalence of NDDs is on the rise due to the aging of the population. The World Health Organization has projected that the number of individuals with NDDs will triple over the next 30 years [[1,](#page-13-0) [2](#page-13-1)].

Although each NDD has distinct characteristics, several common features are observed across these disorders $[3]$ $[3]$. These include pathological protein aggregation, dysfunction of synaptic and neuronal networks, abnor-

mal proteostasis, cytoskeleton abnormality, changes in energy metabolism, defects in DNA and RNA, infamma-tion, and neuronal cell death (Fig. [1](#page-1-0)) $[4]$ $[4]$. The clinical hallmark of NDDs, protein aggregation, aids in the diagnosis of the condition $[5-7]$ $[5-7]$. The spread of neurodegeneration between cells and brain regions is mainly attributed to the misfolding of pathological proteins and the quick spread of their aggregates $[8-11]$ $[8-11]$. Although non-protein NDDs also exist, mature protein aggregates are the primary cause of the illness [[12](#page-13-8)]. Pathologic protein aggregation can occur inside synapses and afect synaptic functions $[13]$ $[13]$. This usually leads to a kind of confusion in the neural network $[14]$ $[14]$. The resultant aberrant neuronal death, impaired energy metabolism, oxidative stress, protein degradation, and cytoskeletal abnormalities

Fig. 1 Hallmarks of NDDs. NDDs are a group of disorders that have several common characteristics. Image adapted from David M Wilson 3rd. Cell. 2023 [\[4](#page-13-3)] with permission from Elsevier Inc. Copyright 2022

are directly linked to synaptic dysfunction [\[15](#page-13-11)[–17](#page-13-12)]. As a result, numerous hallmarks are associated with and involved in NDDs, necessitating the use of multi-targeting therapeutics.

Despite the extensive research efforts dedicated to studying these disorders, current interventional approaches can only ameliorate symptoms rather than halting the progression of NDDs $[18]$ $[18]$. Hence, there is a pressing need for treatments that might signifcantly infuence the progression of neurodegeneration. Multiple pharmaceuticals are now being tested in clinical trials for NDDs; however, a signifcant number of them has been discontinued due to inadequate efectiveness or substantial side efects. Hence, to mitigate the adverse efects of these traditional treatments, innovative therapeutic approaches are needed to achieve successful management of NDDs.

Neuronanotechnology is an innovative method to circumvent obstacles in delivering drugs to the brain. Nanoparticles (NPs) possess distinctive characteristics, including customized size, shape, fexibility, and surface charge, which permit enhancement of therapeutic bioavailability and cellular uptake, reduction of drug resistance, and mitigation of side efects [\[19\]](#page-13-14). Consequently, these materials can serve as suitable vehicles for the diagnosis and treatment of NDDs. Various nanomaterial platforms, such as polymer-based NPs, dendrimers, lipidbased NPs, hydrogels, cyclodextrins, carbon-based NPs, and metal-based NPs, are utilized for therapeutic delivery to the brain $[20]$ $[20]$. There are several advantages associated with this approach. First, it allows for the controlled

drug release at a predetermined rate. Second, NPs may be delivered either locally or systemically. Third, the NPs can remain in the body for a specifed duration. Last, this approach enables targeted delivery of drugs to the precise site of action $[21]$ $[21]$. There are two primary categories of nanostructures that have been developed for nanomedicine applications: biomembrane nanostructures and nonbiomembrane nanostructures (Fig. [2\)](#page-2-0).

Biomembrane nanostructures are formed from biomembranes, including liposomes, cell membranes, extracellular vesicles (EVs), nanostructured lipid carriers (NLCs), and solid lipid nanoparticles (SLNs). These nanostructures possess notable advantages in biocompatibility, multifunctionality, and inherent targeting capability. Furthermore, these nanostructures have the ability to encapsulate both hydrophilic and hydrophobic medications, hence greatly enhancing their versatility [\[22](#page-13-17)]. Nevertheless, the use of biomembrane nanostructures is signifcantly restricted due to the unregulated drug release and inadequate responsiveness. These restrictions can be overcome by modifying the surface and using targeted ligands. Non-biomembrane nanostructures are commonly used in drug carriers, including metal nanostructures, polymers, magnetic NPs, micelles, and hydrogels, due to their distinctive physical and chemical characteristics. The non-biomembrane nanostructures have substantial specifc surface area and their surface morphology can be modified $[23-25]$ $[23-25]$ $[23-25]$. Moreover, they can be fabricated by many techniques, and the metal nanostructures have shown therapeutic efects on animal models of NDDs [[26](#page-13-20), [27\]](#page-13-21). Nevertheless, the

Fig. 2 Advantages and drawbacks of biomembrane nanostructures and non-biomembrane nanostructures. The color green symbolizes the benefts of hybrid nanostructures, while the red color stands for their drawbacks. Image created with Biorender.com

non-biomembrane nanostructures also have side efects. They may induce significant harmful effects on healthy cells [\[28\]](#page-13-22). Furthermore, nanostructures that are not based on biomembranes can be readily eliminated by the reticuloendothelial system (RES), exhibit brief blood circulation durations, and may accumulate in healthy organs such as the liver and spleen, resulting in diminished therapeutic efectiveness [\[29](#page-13-23)]. Consequently, it is imperative to enhance the capacity of non-biomembrane nanostructures to target NDDs and mitigate their side effects $[30]$ $[30]$. This can be achieved by producing hybrid nanostructures.

The hybrid nanostructures have following advantages: (1) long circulation time by reducing nonspecifc phagocytosis and RES clearance [\[31\]](#page-13-25); (2) good biocompatibility achieved through modifcations in the composition, shape, and surface chemical characteristics of the nanostructures [\[32](#page-13-26)]; (3) specific targeting through mechanisms that underlie the inherent targeting of biomembranes or ligand changes; and (4) potent drug-carrying ability for both hydrosoluble and hydrophobic medicines.

There is a scarcity of literature examining the application of hybrid nanostructures for the treatment of NDDs in animal models. In this paper, we provide a comprehensive review of the engineering strategies for hybrid nanostructures, as well as the properties and applications of hybrid nanostructures in the feld of NDD theranostics (Fig. [3\)](#page-3-0).

Fig. 3 An overview of the use of hybrid nanostructures in the detection and management of NDDs, covering drug delivery, engineering approaches for synthesis, and diagnostic imaging. Image created with Biorender.com

Engineering strategies for hybrid nanostructures

The primary goal of hybrid methodologies is to enhance the targeting precision, the biosafety, and the drug delivery capabilities of nanocarriers while preserving the integrity of the blood–brain barrier (BBB) [[33\]](#page-13-27).

Liposomes are the most extensively utilized nanostructures in biomembrane technology. Natural biomembrane-based nanostructures, known as cell membrane-based nanostructures, have a distinctive biomimetic design and can encapsulate various nanocores for the purpose of treating NDDs. EVs are minute membrane vesicles that are released by nearly all live cells. They have the ability to traverse diverse biological obstacles to access specific cells. The SLN is an advanced delivery technology that consists of a solid lipid core and a biocompatible surfactant outer shell. SLN has several distinctive advantages, including a notable drug loading capacity, compact dimensions, increased efective surface area, superior stability compared to alternative lipid carriers, biocompatibility, compatibility with a diverse array of therapeutics, minimal toxicity profle, and enhanced solubility and permeability. The nanostructures SLN and NLC exhibit notable similarities and possess a greater loading capacity [[34](#page-13-28)]. In contrast, metal nanostructures have gained considerable interest in the realm of non-biomembrane nanostructures because of their distinctive characteristics. Metal nanostructures can be disseminated or contained within metal shells and covalently linked to their surfaces, facilitating efective delivery of medicinal medications to the brain. Polymeric NPs have been extensively studied for the purpose of delivering a wide array of treatments to the brain, including tiny hydrophilic molecules, lipophilic medications, proteins, peptides, and several other macromolecules $[35]$ $[35]$. The majority of current magnetic NPs have demonstrated nontoxic properties and exhibited favorable absorption capacities. Additionally, magnetic NPs have the potential to be used as diagnostic instruments [[36\]](#page-13-30). Micelles possess a hydrophilic shell and a hydrophobic core, facilitating the capture of hydrophobic molecules inside the core and hydrophilic components within the shell [[37](#page-13-31)]. Surface modifications with target active ligands can facilitate medication transport to the intended site while decreasing off-site distribution $[38]$ $[38]$. Hydrogels possess numerous advantages, including controlled drug release, degradability, biocompatibility, and stable mechanical strength $[39, 40]$ $[39, 40]$ $[39, 40]$ $[39, 40]$. Therapeutic medications can be released in a continuous and gradual manner throughout the body, resulting in long-term disease therapy [\[41](#page-14-0)].

As stated above, both biomembrane nanostructures and non-biomembrane nanostructures possess inherent limitations. Hybrid nanostructures can address the limitations of both methods by enhancing the permeability of the BBB and the bioavailability of drugs. Various methods can be used to achieve hybridization (Fig. [4\)](#page-5-0).

Co‑extrusion

The most widely used method for producing synthetic liposomes is co-extrusion, usually referred to as physical extrusion $[26]$. The biomembrane and non-biomembrane nanostructure suspensions are combined in a specifc ratio during the co-extrusion process. A porous flter membrane of the extruder with an appropriate size of pores was used to co-extrude the combined suspension several times [\[42](#page-14-1)]. After extrusion, hybrid nanostructures are generated by membrane wrapping of the nano core [[43\]](#page-14-2). In the study by Khongkow et al., for instance, extrusion was utilized to hybridize gold NPs and exosomes, and the resulting hybrid nanostructures showed successful brain targeting [\[44](#page-14-3)]. Moreover, studies have been conducted on hybrid manganese dioxide NPs made from macrophage membranes using the co-extrusion method [[45\]](#page-14-4). The co-extrusion process is straightforward and easy to conduct, and can fabricate hybrid nanostructures of the desired size by modifying the pore size of the flter membrane $[46]$ $[46]$. However, this method causes a significant waste of resources due to the inability to guarantee complete encapsulation of non-biomembrane nanostructures into biomembrane nanostructures.

Ultrasound

Ultrasound treatment of a mixture of non-biomembrane and biomembrane nanostructures leads to the spontaneous generation of hybrid nanostructures [[47](#page-14-6)[–49](#page-14-7)]. Currently, hybrid nanostructures produced using this method have been tested in animal models of NDD. For example, superparamagnetic iron oxide nanoparticles (SPIONs), which are hybrid structures made of exosomes and polymers, have the ability to transport medications to the brain and improve the efectiveness of treatment [[27,](#page-13-21) [50\]](#page-14-8). Compared to the co-extrusion method, the ultrasonic method is more simple to operate and cause less material loss. On the other hand, the membrane coating caused by the ultrasonic approach might not be uniform, and the NPs will be broken during the longterm ultrasonic process [\[31](#page-13-25)].

Direct incubation

Sonication and physical extrusion are time-consuming and require much work. These methods also have a risk of compromising the integrity of biomembrane proteins [[51\]](#page-14-9). Direct incubation is an approach that avoids these drawbacks. Hybrid nanostructures can be fabricated by directly incubating non-biomembrane nanostructures

Fig. 4 Six engineering strategies for hybrid nanostructures, including (1) co-extrusion, (2) ultrasound, (3) direct incubation, (4) direct mixing, (5) thin flm hydration and (6) emulsifcation-solvent evaporation method. Image created with Biorender.com

with cells or biomembrane nanostructures. For instance, Kutchy et al. produced a hybrid nanostructure through direct cell incubation with ultrasminy superparamagnetic iron oxide followed by isolation using a qEV column [\[52](#page-14-10)]. Hydrogels [[53](#page-14-11), [54](#page-14-12)] or gold NPs [\[55](#page-14-13)] can be hybridized with biomembrane nanostructures, extracellular vesicles, or exosomes produced by cells to produce hybrid nanostructures that may be useful for the detection or treatment of NDDs.

Direct mixing

The aforementioned techniques mostly target membrane-containing nanostructures. Hybrid nanostructures cannot be fabricated with non-biomembrane nanostructures using the aforementioned techniques. Under some circumstances, biomembrane nanostructures can be combined directly with non-biomembrane nanostructures to induce hybridization. To treat PD, Uppuluri et al. loaded drug-containing SLNs in a thermoresponsive Methyl Cellulose in situ gel, producing a hybrid structure, which was shown to increase the drug delivery efficiency [\[56](#page-14-14)]. Furthermore, in the study by Adnet et al.,

drug-loaded liposomes were combined with in situ gels to fabricate hybrid nanostructures that could be used to treat AD specifcally [\[57\]](#page-14-15).

Thin flm hydration

One of the conventional techniques for fabricating hybrid nanostructures is the thin flm hydration method. This procedure involves mixing lipids with an organic solvent and removing the organic solvent using rotary evaporation [[58\]](#page-14-16). By hybridizing liposomes and SPIONs, Saesoo et al. developed a hybrid nanostructure that increases the drug delivery efficiency and BBB penetration $[59]$ $[59]$. This technique was further refined by Shi et al. and is known as the lipid thin flm dehydration method [[60\]](#page-14-18). Due to the use of hazardous chemical solvents, the process has potential toxicity, although it is relatively well established and the equipment is straightforward [\[58\]](#page-14-16).

Emulsifcation–solvent evaporation method

The emulsification–solvent evaporation method involves creating an emulsion by stirring or ultrasonic emulsifcation of the aqueous and the organic phases and then

evaporating the organic phase to cause the dispersion phase to encapsulate the medication to create nanostructures [\[61\]](#page-14-19). For instance, Gomes et al. hybridized SLNs and polymers to create hybrid nanostructures that show improved ability to target the brain [[62](#page-14-20)]. Although there are numerous variables infuencing the properties of hybrid nanostructures throughout the fabrication process, this technique has advantages of easy preparation and high reproducibility [\[61\]](#page-14-19).

Other hybridizing methods

In addition to the techniques mentioned above, hybrid nanostructures can also be generated using techniques such as in situ packaging, freeze-thaw/ultrasound, extrusion/ultrasound and stirring, and extrusion/electroporation [[43\]](#page-14-2).

Applications in NDD diagnosis

In the early stages of NDDs, patients are usually asymptomatic, and imaging techniques may provide useful adjunctive information [[63\]](#page-14-21). In advanced stages, the diagnosis of NDDs is largely based on clinical assessment and imaging. Patients usually go to hospitals with symptoms present for a long time. Therefore, early diagnosis and screening for NDDs is crucial. The diagnostic tools for NDDs are very complex, and early stages of various NDDs show many overlapping symptoms that are difficult to distinguish and diferentiate.

NPs are an important tool for molecular imaging involving multiple modes and functions. Superparamagnetic iron oxide, gold NPs/nanorods, manganese oxide (MnO), and quantum dots exhibit distinct characteristics, such as paramagnetism, surface plasmon resonance, superparamagnetism, and photoluminescence. These characteristics render them suitable for imaging, treatment, and drug delivery applications. Techniques used for targeted imaging include magnetic resonance imaging, positron emission tomography, single photon emission computed tomography, photoacoustic imaging, computed tomography (CT), two photon or fuorescent imaging, and ultrasound (Fig. [5](#page-6-0)) [[64\]](#page-14-22).

The main nanostructure hybridization approach investigated for the diagnosis of NDDs is the combination of non-biomembrane nanostructures with exosomes. Exosomes are membrane vesicles that originate from cells and have a size range of 40–100 nm [\[65](#page-14-23)]. Exosomes commonly consist of biological constituents derived from parental cells, including mRNAs, transport proteins, or proteins associated with distinct cellular functions [\[66](#page-14-24)]. Exosomes have the capacity to traverse biological barriers, such as the BBB, due to their signaling molecules, optimum size and membrane coating. Consequently, they may serve as a potent diagnostic tool [[67\]](#page-14-25). However, there are still two major challenges in the feld of exosome research: the ability to prepare exosomes consistently and the efectiveness of tracking exosomes in

Fig. 5 Hybrid nanostructures for the diagnosis of NDDs. Image created with Biorender.com

living organisms. Non-biomembrane nanostructures ofer numerous benefts that can address the limitations of exosomes [\[68\]](#page-14-26). NPs coated with exosomal membranes have been used for imaging and treatment in animal models of NDDs. In a previous study, Au NPs were modifed with neuron-targeted exosomes with the Lamp2b-rabies virus glycoprotein (RVG) and glycosylation-stabilized peptides on their surface [[44](#page-14-3)]. Bioluminescent imaging of the mouse brain revealed that the Au NPs can traverse the BBB and specifcally target brain cells both in vitro and in vivo. Compared with Au NPs coated with unmodifed exosomes, the Au NPs coated with RVGexosomes crossed the BBB more efectively and exhibited greater accumulation in brain cells. This study presents a promising method for overcoming the BBB obstacle and could lead to the development of efficient diagnostic techniques for NDDs [\[44](#page-14-3)]. CT imaging can be used to track exosomes loaded with Au NPs in vivo. For example, exosomes produced from bone marrow mesenchymal stem cells (MSCs) that were loaded with glucose-coated Au NPs were administered in murine models (AD and PD) through the intranasal route. Results showed that the exosomes specifcally targeted and accumulated in pathologically relevant brain regions, with efficacy observed up to 96 h after administration [\[69](#page-14-27)].

Currently, the sole hybrid nanostructure for the diagnostic purposes of NDDs is composed of exosomecoated gold NPs. This hybridization technique has shown potential in enhancing disease diagnosis in experimental models. In future studies, more types of hybrid nanostructures should be explored for their applications in NDD diagnosis.

Applications in NDD treatment

NDDs are difficult to treat. A major obstacle in the management of NDDs is the difficulty of medications to overcome the BBB barrier and selectively target neuronal cells $[26]$ $[26]$. The current therapeutics mostly aim to delay the progression of the diseases, rather than targeting the fundamental etiology [[70\]](#page-14-28). Hybrid nanostructures, which involve the combination of biomembrane nanostructures with non-biomembrane nanostructures, show a substantial increase in BBB permeability and precise targeting of lesion sites in NDDs. This leads to major improvements in both diagnostic accuracy and therapeutic efficacy.

NDDs exhibit some common pathological characteristics, including neuronal loss, aberrant accumulation of protein aggregates, and persistent infammation in specific brain areas $[71]$ $[71]$ $[71]$. Therefore, hybrid nanostructures are able to treat a wide range of NDDs by targeting these common pathological features. Li et al. designed macrophage-disguised $MnO₂$ NPs loaded with fingolimod, and found that these NPs could consume reactive oxygen species (ROS) and produce oxygen (O_2) . Furthermore, the NPs can counteract the proinfammatory milieu by facilitating the phenotypic transformation of microglia via various signaling pathways, hence enhancing the protective effects on injured neurons $[45]$ $[45]$. Feng et al. developed a mesoporous Prussian blue nanozyme (MPBzyme@NCM) with a neutrophil-like cell-membrane (NCM) coating. They found that the MPBzyme $@$ NCM can traverse the BBB and scavenge ROS when administered intravenously. Compared with both nondifferentiated HL-60 cell membrane coating and noncoated condition, the NCM coating resulted in sustained accumulation of MPBzyme@NCM in the damaged brain [[72\]](#page-14-30).

Hydrogels are a three-dimensional cross-linked polymeric network that possesses a very hydrophilic structure [[41\]](#page-14-0). Hydrogels have been extensively employed in the therapeutics of diverse diseases owing to their advantageous properties, including biodegradability, biocompatibility, robust mechanical integrity, and controlled release of drugs [[39](#page-13-33), [40\]](#page-13-34). Long-term neuroinfammation poses a signifcant barrier to neurological recovery in the context of neurodegenerative disorders. Zhang et al. developed a method to combine hydrogels with exosomes to deliver exosomes derived from interleukin-1β-stimulated bone marrow stromal cells (βExos) in a sustainable manner. This approach enhances exosome production and anti-infammatory capabilities, leading to the inhibition of neuroinfammation and promotion of neurological recovery. Following in situ injection into the brain injury site, the released βExos effectively regulated neuroinflammation, resulting in considerable reductions of glial scar formation and neuronal loss [[54](#page-14-12)]. Similarly, exosomes can be combined with SPIONs to create hybrid nanostructures that specifcally target and treat animal models of NDDs. In the study by Peng et al. [\[27\]](#page-13-21), the ROS-responsive polymer poly(propylene sulfde)-polyethylene glycol (PPS-PEG) were loaded with SPIONs and hydrophobic curcumin to form the micellar core ($PP@Cur$). The outer layer (PR-EXO) was formed by embedding SA-P (stearic acid-RQIKIWFQNRRMKWKK) and SA-RVG (stearylamine-YTIWMPENPRPGTPCDIFTNSRGKRASNG) into the phospholipid bilayer of exosomes. The self-oriented nanocarrier (PR-EXO/PP@Cur) was fabricated by mixing the outer layer and the core and passing them through an extruder. After intranasal treatment, brain accumulation of PR-EXO/PP@Cur was observed within 12 h of nasal treatment (Fig. [6](#page-8-0)a, b). Furthermore, curcumin accumulation in the brain was detected ex vivo at 6 h after nasal administration of PR-EXO/PP@Cur (Fig. [6c](#page-8-0)). Compared to other treatment groups of MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) PD mice, three weeks of PR-EXOs/PP@Cur treatment led to signifcantly decreased microglial infltration in PD

mice (Fig. [6](#page-8-0)d). Additionally, the administration of PR-EXOs/PP@Cur restored microglial morphology (Fig. [6e](#page-8-0)), decreased the concentrations of proinfammatory factors, such as TNF-α, interleukin (IL)-1β, and IL-6, while increasing the anti-infammatory factor IL-10 in PD mice (Fig. [6f](#page-8-0)). Among the NPs tested, mice with PR-EXO/PP@ Cur treatment exhibited the most pronounced enhancement in movement behavior and coordination ability (Fig. [6g](#page-8-0), h) [[27](#page-13-21)].

There are also studies focusing on the effects of hybrid nanostructures on specifc pathological symptoms of a particular NDD [[27,](#page-13-21) [50](#page-14-8), [73](#page-14-31), [74](#page-14-32)]. In the following, we will focus on three diseases: AD, PD, and MS.

AD

AD is the prevailing type of dementia globally, with an estimated prevalence ranging from 50% to 80% of cases [[75\]](#page-14-33). The etiology of AD is intricate, and its pathophysiology remains elusive. The main pathological characteristics include atypical accumulation of beta-amyloid protein (Aβ), aberrant phosphorylation of tau protein, and infammation of the brain system, among other potential factors [\[76](#page-14-34)]. Currently, there are still few available therapies for AD.

One of the main treatments for AD that is being developed clinically is passive immunotherapy. Aβ peptide deposition is thought to be the primary initiator of neurodegenerative processes in AD [\[77](#page-14-35)]. Recent research shows that monoclonal antibodies can slow early AD progression and stimulate Aβ elimination from the brain. The antibodies bapineuzumab, solanezumab, crenezumab, gantenerumab, aducanumab, lecanemab, and donanemab have progressed the furthest in clinical development. The antibodies lecanemab, donanemab, and aducanumab can remove > 60% of $\text{A}\beta$ deposits after 18 months of treatment, and have demonstrated efficacy in slowing clinical decline [\[78](#page-14-36)]. Lecanemab, however, was the frst to obtain complete Food and Drug Administration (FDA) approval. Furthermore, only people with mild to moderate AD might beneft from monoclonal antibody medications in terms of cognitive performance. During treatment, doctors must be aware of the possible side efects including cerebral edema [\[79\]](#page-14-37). Researchers have investigated the potential therapeutic benefts of hybrid nanostructures in AD.

Cell membrane-coated NPs have been extensively studied as a potential solution to address this issue. This approach enhances the overall efficiency of nano-drug delivery systems [[43\]](#page-14-2). Tang et al. fabricated macrophage membrane-encapsulated, rapamycin-loaded 4-(hydroxymethyl) phenylboronic acid pinacol ester NPs that are responsive to ROS. They found that the biomimetic NPs could specifcally target the ROS region and release phar-maceuticals, with enhanced efficacy in drug delivery [\[80](#page-14-38)]. While their study tested application of the biomimetic NPs in atherosclerosis, these results also imply potential for AD as ROS also increase in AD. Thus, studies of hybrid NPs for the treatment of AD are anticipated.

Due to the biocompatibility and slow release properties of hydrogels, Cunha et al. developed in situ thermosensitive nasal gels with hybridized NLCs for direct delivery of rivastigmine to the brain. This nanosystem showed adequate nasal mucoadhesion and sustained drug release, providing a promising new treatment strategy for advancing the management of AD [\[81\]](#page-14-39). Adnet et al. integrated the liposome and hydrogel methodologies to create a composite nanosystem for nasal AD treatment. The results showed increased drug residence time in the nasal cavity and regulated release of the drug [\[57](#page-14-15)]. In a recent study examining the capacity of NPs and exosomes to improve neurological illnesses, copper sulfde NPs and exosomes produced from MSCs were administered together in rats with cadmium (Cd)-induced neurological disorders. The combined treatment improved total antioxidant status, increased BDNF and NGF, and showed anti-infammatory efects. Histological assessments demonstrated that the combined treatment reduced the deleterious impacts of Cd on brain tissue [[82](#page-14-40)].

In summary, hybrid nanostructures increase drug loading capacity, extend drug circulation time, improve drug

(See fgure on next page.)

Fig. 6 Biodistribution of drugs in the brain and therapeutic efects in vivo after nasal administration. **a** Fluorescence intensity of Cy7 in the brains of PD mice. **b** The mean fuorescence intensity of Cy7 in the mouse brain regions quantifed from **a**. **c** Ex vivo biodistribution of diferent NPs in PD mouse brains after 6 h of nasal administration. Labels I–VI in panels **a** and **c** represent PBS, PP@Cur, EXO/PP@Cur, P-EXO/PP@Cur, R-EXO/ PP@Cur, and PR-EXO/PP@Cur, respectively. Data are presented as the mean±SD (*n*=3). **d** Immunofuorescence staining of Iba-1 in the substantia nigra of wild-type and PD mice treated with diferent NPs. **e** Magnifed images of microglia in wild-type mice (I), PD mice (II), and PD mice treated with the PR-EXO/PP@Cur nanocarriers (VI). **f** Serum concentrations of TNF-α, IL-1β, IL-6, and IL-10 in wild-type and PD mice treated with diferent NPs (*n*=4). **g** Trajectories of wild-type and PD mice treated with diferent NPs in the open-feld test. Labels I–VII in panels **d**, **e** and **g** represent Wild-type, MPTP-PD, PP@Cur, EXO/PP@Cur, PR-EXO133b–/PP@Cur, PR-EXO/PP@Cur, and PR-EXO/PP@Cur, respectively. **h** Total distance travelled, speed, and total resting time of wild-type and PD mice treated with diferent NPs in the open-feld test. Scoring of wild-type and PD mice treated with diferent NPs in the pole-climbing test. Data are presented as the mean±SD (*n*=5). Student's *t* test: **P*<0.05; ***P*<0.01; ****P*<0.001; ns, not signifcantly diferent. Adapted from Peng H, et al. ACS Nano. 2022 [[27\]](#page-13-21) with permission from the American Chemical Society. Copyright 2022

Fig. 6 (See legend on previous page.)

Table 1 Summary of studies on hybrid nanostructures based on biomembrane and non-biomembrane nanostructures for the treatment of AD

Nanomedicine	Biomembrane nanostructure	Non- biomembrane nanostructure	Method of administration Therapeutic effects		Research stage	Reference
Liposomes of API	Liposomes	Hydrogel	Intranasal administration	Prolong API release and increase API residence time	In vitro	$[57]$
MSC-Exo+CuSNPs MSC-Exo		CuSNPs	Oral administration	Ameliorate the oxidative stress and inflammatory mark- ers; protect neurons	In vivo	[82]
In situ gel of rivastigmine- loaded NLC	NLC	In situ gel	Intranasal administration	Increase the viscosity and the mucoadhesion; show prolonged drug release	In vitro	[81]

API, Active Pharmaceutical Ingredient; CuSNP, copper sulfde nanoparticle; MSC-Exo, exosomes derived from mesenchymal stem cells

targeting, and achieve therapeutic efects in AD treatment. Nevertheless, this strategy has not been extensively studied for AD treatment, and most of them are still in the preclinical stage (Table [1\)](#page-10-0).

PD

PD remains untreatable. Current therapeutic strategies primarily aim to enhance dopaminergic signaling. These therapies include levodopa (*L*-dopa) that inhibits DOPA decarboxylase, inhibitors of catechol-Omethyltransferase, dopamine agonists, and inhibitors of the enzyme monoamine oxidase type B [[83\]](#page-14-41). Regrettably, the aforementioned treatments solely alleviate symptoms without impeding the clinical and pathological progression of PD [[84](#page-14-42)].

The advancement of hybridization technology offers a viable approach for the management of PD. Liu et al. fabricated a RVG-exosome (REXO)-coated genechem nanocomplex REXO-C/ANP/S, by hybridising exosome and polymer. This nanocomplex effectively concentrates medications at the action site of a target cell. At the cellular and animal levels, the functions of REXO-C/ANP/S in clearing α -syn aggregates through the BBB and facilitating membrane fusion have been verified. This delivery system achieved efficient administration of siRNA and chemical drugs and reduced α-syn aggregates in diseased dopaminergic neurons [[50](#page-14-8)]. In another study, to ensure effective transportation of piribedil to the brain, the SLNs were encapsulated in a thermoresponsive methyl cellulose in situ gel (PBD-SLN-ISG) to delay mucociliary clearance after oral administration in rats. Pharmacokinetic experiments conducted in living organisms demonstrated that compared with simple intranasal suspensions of PBD (PBD-Susp), PBD-SLN-ISG resulted in a 4-fold increase in the PBD $(AUC)_{brain}$ and a 2.3-fold decrease in PBD $(C_{\text{max}})_{\text{plasma}}$. In addition, PBD-Susp showed limited direct absorption from the nose to the brain, while the optimized PBD-SLN-ISG demonstrated efficient direct nose-to-brain absorption [\[56\]](#page-14-14).

MS

MS is an immune-mediated infammatory disease of the the CNS, characterized by the presence of infammatory lesions, demyelinating plaques, and irreversible damage to axons as the disease progresses [\[85](#page-14-43)]. Relapsing– remitting MS is the most common MS phenotype and is characterized by recurring relapses and remissions of neurological symptoms in affected individuals $[86]$ $[86]$. The aim of MS treatment is to minimize relapses and slow the course of the disease [\[87](#page-15-1)]. Disease-modifying treatments (DMTs) are commonly employed in long-term management to provide therapeutic advantages by inhibiting or regulating immune function and infammatory responses [[87,](#page-15-1) [88](#page-15-2)]. Nevertheless, the existing DMTs currently demonstrate restricted efficacy in arresting the course of MS [[89\]](#page-15-3).

With the advent of stem cell therapies, Ferreira et al. employed hydrogels and liposomes as a means to fabricate hybrid nanostructures for the purpose of delivering MSCs in the context of MS treatment. The engineered hydrogel carrying MSCs was administered intracerebroventricularly in an experimental autoimmune encephalomyelitis rat model. Compared with cell suspensions, the formulated solution was more efective in decreasing the severity of the disease and achieving maximum clinical score. Therefore, the hybrid nanostructures may provide a promising avenue for addressing the challenges

Fig. 7 Challenges and perspectives of hybrid nanostructures for AD treatment. Image created with Biorender.com

associated with existing therapeutic approaches in the treatment of MS [[90](#page-15-4)].

Others

NDDs include many diseases. We searched literature in PubMed and the Web of Science using the following keywords: (((((((((((Hybrid Nanostructures[Title]) OR (liposome[Title])) OR (cell membrane[Title])) OR (extracellular vesicle[Title])) OR (nanostructured lipid carriers[Title])) OR (solid lipid nanoparticles[Title])) OR (metal nanostructure[Title])) OR (polymer[Title])) OR (magnetic nanoparticle[Title])) OR (micelle[Title])) OR (hydrogel[Title])) AND (((((((((Neurodegenerative diseases[Title]) OR (Alzheimer disease[Title])) OR (Parkinson disease[Title])) OR (primary tauopathies[Title]))

OR (frontotemporal dementia[Title])) OR (amyotrophic lateral sclerosis[Title])) OR (synucleinopathies[Title])) OR (Huntington disease[Title])) OR (related polyglutamine (polyQ) diseases[Title])). However, we found no studies using hybrid nanostructures as carriers for the treatment of other NDDs.

Cholesterol production is markedly reduced in animal models of HD, and this is closely correlated with both synaptic dysfunction and cognitive impairment [\[91](#page-15-5)]. One potential HD treatment is to increase the amount of cholesterol through efective delivery to the brain [[92\]](#page-15-6). In the study by Tosi et al., polylactic-co-glycolic acid (PLGA) and cholesterol were mixed to generate a novel NP that could transfer peripheral cholesterol across the BBB. The conjugated g7-glycopeptide (H₂N-Glyl-Phe-D-T

hr-Gly-*L*-Phe-*L*-Leu-*L*-Ser[*O*-β-*D*-glucose]-CONH2) on the surface of the NPs promotes the passage of particles across the BBB in a non-invasive manner by producing BBB membrane curvature and subsequent endocytosis [[93\]](#page-15-7). In a more recent study by the same research group, the cholesterol and PLGA hybrid NPs modifed with g7 (g7-PLGA-cholesterol) were found to be taken up by neurons mainly through the clathrin-dependent mechanism, and the g7-glycopeptide had a major efect on the interaction between the NPs and the cells. However, if the passage through the BBB is mediated by receptors, the amount of NPs that could reach the brain would be lower due to the potential saturation of these receptors. In conclusion, this gives us optimism for HD therapies and the use of hybrid nanostructures in the treatment of NDDs, even though this NP is not a hybridization of biomembrane and non-biomembrane nanostructures [[94\]](#page-15-8). Research on the benefts of hybrid nanostructures in medicine delivery is currently lacking. Next, it is imperative to achieve accurate treatment outcomes by regulating and targeting the pathological environment in a targeted manner.

Conclusions and prospects

NDD diagnosis and treatment are challenging. Many patients missed timely treatment due to delayed diagnosis. In addition, the longer the treatment duration is, the more detrimental the treatment outcome will be [\[95](#page-15-9)]. The nanotechnology holds great potential in the field of NDDs. Nanostructures can efectively transport pharmaceuticals to the brain while preserving the integrity of the BBB, hence serving as a therapeutic platform. This article describes various hybrid approaches involving biomembrane and non-biomembrane nanostructures. These methodologies possess distinct merits and shortcomings. Hybrid nanostructures hold signifcant value in the feld of diagnosis and treatment of NDDs. Currently, the primary focus of research on hybrid nanostructures for the treatment of NDDs lies in the areas of AD and PD. By employing certain modifcation techniques [\[26](#page-13-20)], hybrid biomembrane nanostructures and non-biomembrane nanostructures have demonstrated signifcant therapeutic efficacy in basic research on NDDs.

However, the hybrid technology is primarily confned to experimental research and has yet to reach clinical implementation. To achieve the desired performance, several modifcation techniques are needed [\[33\]](#page-13-27), such as genetic engineering, covalent coupling, and non-covalent modifications. The modification approaches play a crucial role in enhancing the BBB penetration and the braintargeting capabilities of hybrid nanostructures.

Despite the promising advancements in hybrid nanostructures, there are several obstacles that must be addressed (Fig. [7\)](#page-11-0). (1) The features of biological membrane nanostructures, such as cell membrane and exosomes, vary depending on the origin and the constituent membrane elements. Hybrid techniques that combine the distinct characteristics of diferent biological membranes are recommended. (2) Currently, studies of hybrid nanostructures for the treatment of NDDs in animal models are primarily focused on AD and PD. However, it is important to conduct further research on the treatment of other NDD diseases. (3) Many diseases are caused by more than one single pathological factor. Employing multi-target treatment strategies may signifcantly enhance therapeutic outcomes. (4) Proteins present on the biomembrane have the potential to elicit immunological responses. Biomaterials are present in non-biomembrane nanostructures. Further studies are warranted to examine the immunogenicity and potential side efects of biomembrane nanostructures before their clinical translation. (5) Additionally, standardization of the separation method for biomembrane nanostructures and the hybrid technology for fabricating hybrid nanostructures remains unresolved. Addressing these concerns may pave the way for therapeutic applications of hybrid nanostructures.

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Consent for publication

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References

- 1. Asefy Z, Hoseinnejhad S, Ceferov Z. Nanoparticles approaches in neurodegenerative diseases diagnosis and treatment. Neurol Sci. 2021;42:2653–60.
- 2. Chang CJ, Cravatt BF, Johnson DS, Lim MH. Molecular medicine and neurodegenerative diseases. Chem Soc Rev. 2014;43:6668–71.
- 3. Palanisamy CP, Pei J, Alugoju P, Anthikapalli NVA, Jayaraman S, Veeraraghavan VP, et al. New strategies of neurodegenerative disease treatment with extracellular vesicles (EVs) derived from mesenchymal stem cells (MSCs). Theranostics. 2023;13:4138–65.
- 4. Wilson DM 3rd, Cookson MR, Van Den Bosch L, Zetterberg H, Holtzman DM, Dewachter I. Hallmarks of neurodegenerative diseases. Cell. 2023;186:693–714.
- 5. Erkkinen MG, Kim MO, Geschwind MD. Clinical neurology and epidemiology of the major neurodegenerative diseases. Cold Spring Harb Perspect Biol. 2018;10.
- 6. Dugger BN, Dickson DW. Pathology of neurodegenerative diseases. Cold Spring Harb Perspect Biol. 2017;9.
- 7. Prusiner SB. Shattuck lecture–neurodegenerative diseases and prions. N Engl J Med. 2001;344:1516–26.
- 8. Flach M, Leu C, Martinisi A, Skachokova Z, Frank S, Tolnay M, et al. Transseeding of Alzheimer-related tau protein by a yeast prion. Alzheimers Dement. 2022;18:2481–92.
- 9. Soto C, Satani N. The intricate mechanisms of neurodegeneration in prion diseases. Trends Mol Med. 2011;17:14–24.
- 10. Scheckel C, Aguzzi A. Prions, prionoids and protein misfolding disorders. Nat Rev Genet. 2018;19:405–18.
- 11. Prusiner SB. Molecular biology of prion diseases. Science. 1991;252:1515–22.
- 12. Kayed R, Head E, Thompson JL, McIntire TM, Milton SC, Cotman CW, et al. Common structure of soluble amyloid oligomers implies common mechanism of pathogenesis. Science. 2003;300:486–9.
- 13. Spires-Jones TL, Hyman BT. The intersection of amyloid beta and tau at synapses in Alzheimer's disease. Neuron. 2014;82:756–71.
- 14. Seeley WW, Crawford RK, Zhou J, Miller BL, Greicius MD. Neurodegenerative diseases target large-scale human brain networks. Neuron. 2009;62:42–52.
- 15. Sudhof TC. The presynaptic active zone. Neuron. 2012;75:11–25.
- 16. Li S, Sheng ZH. Energy matters: presynaptic metabolism and the maintenance of synaptic transmission. Nat Rev Neurosci. 2022;23:4–22.
- 17. Cunnane SC, Trushina E, Morland C, Prigione A, Casadesus G, Andrews ZB, et al. Brain energy rescue: an emerging therapeutic concept

for neurodegenerative disorders of ageing. Nat Rev Drug Discov. 2020;19:609–33.

- 18. Armstrong MJ, Okun MS. Diagnosis and treatment of Parkinson disease: a review. JAMA. 2020;323:548–60.
- 19. Esmaeili Y, Yarjanli Z, Pakniya F, Bidram E, Los MJ, Eshraghi M, et al. Targeting autophagy, oxidative stress, and ER stress for neurodegenerative disease treatment. J Control Release. 2022;345:147–75.
- 20. Liaw K, Zhang Z, Kannan S. Neuronanotechnology for brain regeneration. Adv Drug Deliv Rev. 2019;148:3–18.
- 21. Bassyouni F, ElHalwany N, Abdel Rehim M, Neyfeh M. Advances and new technologies applied in controlled drug delivery system. Res Chem Intermed. 2013;41:2165–200.
- 22. Campani V, Giarra S, De Rosa G. Lipid-based core-shell nanoparticles: evolution and potentialities in drug delivery. OpenNano. 2018;3:5–17.
- 23. Vimbela GV, Ngo SM, Fraze C, Yang L, Stout DA. Antibacterial properties and toxicity from metallic nanomaterials. Int J Nanomed. 2017;12:3941–65.
- 24. Zhao W, Li A, Zhang A, Zheng Y, Liu J. Recent advances in functionalpolymer-decorated transition-metal nanomaterials for bioimaging and cancer therapy. ChemMedChem. 2018;13:2134–49.
- 25. Minakshi P, Mohan H, Manjeet, Ravina, Brar B, Shafq M, et al. Organic polymer and metal nano-particle based composites for improvement of the analytical performance of electrochemical biosensors. Curr Top Med Chem. 2020;20:1029–41.
- 26. Lopes D, Lopes J, Pereira-Silva M, Peixoto D, Rabiee N, Veiga F, et al. Bioengineered exosomal-membrane-camouflaged abiotic nanocarriers: neurodegenerative diseases, tissue engineering and regenerative medicine. Mil Med Res. 2023;10:19.
- 27. Peng H, Li Y, Ji W, Zhao R, Lu Z, Shen J, et al. Intranasal administration of self-oriented nanocarriers based on therapeutic exosomes for synergistic treatment of Parkinson's disease. ACS Nano. 2022;16:869–84.
- 28. Lopez-Chaves C, Soto-Alvaredo J, Montes-Bayon M, Bettmer J, Llopis J, Sanchez-Gonzalez C. Gold nanoparticles: distribution, bioaccumulation and toxicity. In vitro and in vivo studies. Nanomedicine. 2018;14:1–12.
- 29. Almeida JP, Chen AL, Foster A, Drezek R. In vivo biodistribution of nanoparticles. Nanomedicine (Lond). 2011;6:815–35.
- 30. Neha D, Momin M, Khan T, Gharat S, Ningthoujam RS, Omri A. Metallic nanoparticles as drug delivery system for the treatment of cancer. Expert Opin Drug Deliv. 2021;18:1261–90.
- 31. Luk BT, Zhang L. Cell membrane-camoufaged nanoparticles for drug delivery. J Control Release. 2015;220:600–7.
- 32. Amini SM, Kharrazi S, Rezayat SM, Gilani K. Radiofrequency electric feld hyperthermia with gold nanostructures: role of particle shape and surface chemistry. Artif Cells Nanomed Biotechnol. 2018;46:1452–62.
- 33. Liang Y, Iqbal Z, Lu J, Wang J, Zhang H, Chen X, et al. Cell-derived nanovesicle-mediated drug delivery to the brain: Principles and strategies for vesicle engineering. Mol Ther. 2023;31:1207–24.
- 34. Agrawal M, Prathyusha E, Ahmed H, Dubey SK, Kesharwani P, Singhvi G, et al. Biomaterials in treatment of Alzheimer's disease. Neurochem Int. 2021;145: 105008.
- 35. Ferreira NN, Granja S, Boni FI, Prezotti FG, Ferreira LMB, Cury BSF, et al. Modulating chitosan-PLGA nanoparticle properties to design a codelivery platform for glioblastoma therapy intended for nose-to-brain route. Drug Deliv Transl Res. 2020;10:1729–47.
- 36. Liu XG, Zhang L, Lu S, Liu DQ, Zhang LX, Yu XL, et al. Multifunctional superparamagnetic iron oxide nanoparticles conjugated with abeta oligomer-specifc SCFV antibody and class a scavenger receptor activator show early diagnostic potentials for Alzheimer's disease. Int J Nanomedicine. 2020;15:4919–32.
- 37. Shah S, Leon L. Structural dynamics, phase behavior, and applications of polyelectrolyte complex micelles. Curr Opin Colloid In. 2021;53.
- 38. Agwa MM, Abdelmonsif DA, Khattab SN, Sabra S. Self- assembled lactoferrin-conjugated linoleic acid micelles as an orally active targeted nanoplatform for Alzheimer's disease. Int J Biol Macromol. 2020;162:246–61.
- 39. Rajkovic O, Potjewyd G, Pinteaux E. Regenerative medicine therapies for targeting neuroinfammation after stroke. Front Neurol. 2018;9:734.
- 40. Ojeda-Hernandez DD, Canales-Aguirre AA, Matias-Guiu J, Gomez-Pinedo U, Mateos-Diaz JC. Potential of chitosan and its derivatives for biomedical applications in the central nervous system. Front Bioeng Biotechnol. 2020;8:389.
- 41. Gong B, Cheng W, Ji W, Chen X, Chu J, Liang W, et al. Hydrogel: a promising new technique for treating Alzheimer's disease. J Transl Int Med. 2022;10:15–7.
- 42. Long Y, Xiang Y, Liu S, Zhang Y, Wan J, Ci Z, et al. Macrophage membrane modifed baicalin liposomes improve brain targeting for alleviating cerebral ischemia reperfusion injury. Nanomedicine. 2022;43: 102547.
- 43. Zhong X, Na Y, Yin S, Yan C, Gu J, Zhang N, et al. Cell membrane biomimetic nanoparticles with potential in treatment of Alzheimer's disease. Molecules. 2023;28.
- 44. Khongkow M, Yata T, Boonrungsiman S, Ruktanonchai UR, Graham D, Namdee K. Surface modifcation of gold nanoparticles with neurontargeted exosome for enhanced blood-brain barrier penetration. Sci Rep. 2019;9:8278.
- 45. Li C, Zhao Z, Luo Y, Ning T, Liu P, Chen Q, et al. Macrophage-disguised manganese dioxide nanoparticles for neuroprotection by reducing oxidative stress and modulating infammatory microenvironment in acute ischemic stroke. Adv Sci (Weinh). 2021;8: e2101526.
- 46. Rahman MA, Wang J, Zhang C, Olah A, Baer E. Novel micro-/nanoporous cellular membranes by forced assembly co-extrusion technology. Eur Polymer J. 2016;83:99–113.
- 47. He W, Frueh J, Wu Z, He Q. Leucocyte membrane-coated Janus microcapsules for enhanced photothermal cancer treatment. Langmuir. 2016;32:3637–44.
- 48. Liu Q, Fan T, Zheng Y, Yang SL, Yu Z, Duo Y, et al. Immunogenic exosome-encapsulated black phosphorus nanoparticles as an efective anticancer photo-nanovaccine. Nanoscale. 2020;12:19939–52.
- 49. Xiong F, Ling X, Chen X, Chen J, Tan J, Cao W, et al. Pursuing specifc chemotherapy of orthotopic breast cancer with lung metastasis from docking nanoparticles driven by bioinspired exosomes. Nano Lett. 2019;19:3256–66.
- 50. Liu L, Li Y, Peng H, Liu R, Ji W, Shi Z, et al. Targeted exosome coating gene-chem nanocomplex as "nanoscavenger" for clearing alpha-synuclein and immune activation of Parkinson's disease. Sci Adv. 2020;6.
- 51. Lu M, Huang Y. Bioinspired exosome-like therapeutics and delivery nanoplatforms. Biomaterials. 2020;242: 119925.
- 52. Kutchy NA, Ma R, Liu Y, Buch S, Hu G. Extracellular vesicle-mediated delivery of ultrasmall superparamagnetic iron oxide nanoparticles to mice brain. Front Pharmacol. 2022;13: 819516.
- 53. Jiang Y, Wang R, Wang C, Guo Y, Xu T, Zhang Z, et al. Brain microenvironment responsive and pro-angiogenic extracellular vesicle-hydrogel for promoting neurobehavioral recovery in type 2 diabetic mice after stroke. Adv Healthc Mater. 2022;11: e2201150.
- 54. Zhang M, Zhang R, Chen H, Zhang X, Zhang Y, Liu H, et al. Injectable supramolecular hybrid hydrogel delivers IL-1beta-stimulated exosomes to target neuroinfammation. ACS Appl Mater Interfaces. 2023;15:6486–98.
- 55. Betzer O, Perets N, Angel A, Motiei M, Sadan T, Yadid G, et al. In vivo neuroimaging of exosomes using gold nanoparticles. ACS Nano. 2017;11:10883–93.
- 56. Uppuluri CT, Ravi PR, Dalvi AV. Design, optimization and pharmacokinetic evaluation of Piribedil loaded solid lipid nanoparticles dispersed in nasal in situ gelling system for efective management of Parkinson's disease. Int J Pharm. 2021;606: 120881.
- 57. Adnet T, Groo AC, Picard C, Davis A, Corvaisier S, Since M, et al. Pharmacotechnical development of a nasal drug delivery composite nanosystem intended for Alzheimer's disease treatment. Pharmaceutics. 2020;12.
- 58. Shah S, Dhawan V, Holm R, Nagarsenker MS, Perrie Y. Liposomes: advancements and innovation in the manufacturing process. Adv Drug Deliv Rev. 2020;154–155:102–22.
- 59. Saesoo S, Sathornsumetee S, Anekwiang P, Treetidnipa C, Thuwajit P, Bunthot S, et al. Characterization of liposome-containing SPIONs conjugated with anti-CD20 developed as a novel theranostic agent for central nervous system lymphoma. Colloids Surf B Biointerfaces. 2018;161:497–507.
- 60. Shi D, Mi G, Shen Y, Webster TJ. Glioma-targeted dual functionalized thermosensitive Ferri-liposomes for drug delivery through an in vitro blood-brain barrier. Nanoscale. 2019;11:15057–71.
- 61. Pooja D, Tunki L, Kulhari H, Reddy BB, Sistla R. Optimization of solid lipid nanoparticles prepared by a single emulsifcation-solvent evaporation method. Data Brief. 2016;6:15–9.
- 62. Gomes MJ, Fernandes C, Martins S, Borges F, Sarmento B. Tailoring lipid and polymeric nanoparticles as siRNA carriers towards the blood-brain barrier - from targeting to safe administration. J Neuroimmune Pharmacol. 2017;12:107–19.
- 63. Stoessl AJ. Neuroimaging in the early diagnosis of neurodegenerative disease. Transl Neurodegener. 2012;1:5.
- 64. Padmanabhan P, Kumar A, Kumar S, Chaudhary RK, Gulyas B. Nanoparticles in practice for molecular-imaging applications: an overview. Acta Biomater. 2016;41:1–16.
- 65. Thery C, Zitvogel L, Amigorena S. Exosomes: composition, biogenesis and function. Nat Rev Immunol. 2002;2:569–79.
- 66. Hessvik NP, Llorente A. Current knowledge on exosome biogenesis and release. Cell Mol Life Sci. 2018;75:193–208.
- 67. Elliott RO, He M. Unlocking the power of exosomes for crossing biological barriers in drug delivery. Pharmaceutics. 2021;13.
- 68. Barjesteh T, Mansur S, Bao Y. Inorganic nanoparticle-loaded exosomes for biomedical applications. Molecules. 2021;26.
- 69. Perets N, Betzer O, Shapira R, Brenstein S, Angel A, Sadan T, et al. Golden exosomes selectively target brain pathologies in neurodegenerative and neurodevelopmental disorders. Nano Lett. 2019;19:3422–31.
- 70. Wang Z, Gonzalez KM, Cordova LE, Lu J. Nanotechnology-empowered therapeutics targeting neurodegenerative diseases. Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2023;15: e1907.
- 71. Lee HJ, Yoon YS, Lee SJ. Molecular mechanisms of cellular senescence in neurodegenerative diseases. J Mol Biol. 2023;435: 168114.
- 72. Feng L, Dou C, Xia Y, Li B, Zhao M, Yu P, et al. Neutrophil-like cell-membrane-coated nanozyme therapy for ischemic brain damage and longterm neurological functional recovery. ACS Nano. 2021;15:2263–80.
- 73. Han Y, Chu X, Cui L, Fu S, Gao C, Li Y, et al. Neuronal mitochondriatargeted therapy for Alzheimer's disease by systemic delivery of resveratrol using dual-modifed novel biomimetic nanosystems. Drug Deliv. 2020;27:502–18.
- 74. Han Y, Gao C, Wang H, Sun J, Liang M, Feng Y, et al. Macrophage membrane-coated nanocarriers Co-Modifed by RVG29 and TPP improve brain neuronal mitochondria-targeting and therapeutic efficacy in Alzheimer's disease mice. Bioact Mater. 2021;6:529–42.
- 75. Collaborators GBDD. Global, regional, and national burden of Alzheimer's disease and other dementias, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. Lancet Neurol. 2019;18:88–106.
- 76. Peng Y, Jin H, Xue YH, Chen Q, Yao SY, Du MQ, et al. Current and future therapeutic strategies for Alzheimer's disease: an overview of drug development bottlenecks. Front Aging Neurosci. 2023;15:1206572.
- 77. Guo X, Yan L, Zhang D, Zhao Y. Passive immunotherapy for Alzheimer's disease. Ageing Res Rev. 2024;94: 102192.
- 78. Jucker M, Walker LC. Alzheimer's disease: from immunotherapy to immunoprevention. Cell. 2023;186:4260–70.
- 79. van Dyck CH, Swanson CJ, Aisen P, Bateman RJ, Chen C, Gee M, et al. Lecanemab in early Alzheimer's disease. N Engl J Med. 2023;388:9–21.
- 80. Tang D, Wang Y, Wijaya A, Liu B, Maruf A, Wang J, et al. ROS-responsive biomimetic nanoparticles for potential application in targeted antiatherosclerosis. Regen Biomater. 2021;8:rbab033.
- 81. Cunha S, Swedrowska M, Bellahnid Y, Xu Z, Sousa Lobo JM, Forbes B, et al. Thermosensitive in situ hydrogels of rivastigmine-loaded lipid-based nanosystems for nose-to-brain delivery: characterisation, biocompatibility, and drug deposition studies. Int J Pharm. 2022;620: 121720.
- 82. Zaazaa AM, Abd El-Motelp BA, Ali NA, Youssef AM, Sayed MA, Mohamed SH. Stem cell-derived exosomes and copper sulfide nanoparticles attenuate the progression of neurodegenerative disorders induced by cadmium in rats. Heliyon. 2022;8: e08622.
- 83. Kakkar AK, Singh H, Medhi B. Old wines in new bottles: repurposing opportunities for Parkinson's disease. Eur J Pharmacol. 2018;830:115–27.
- 84. Agostini F, Masato A, Bubacco L, Bisaglia M. Metformin repurposing for parkinson disease therapy: opportunities and challenges. Int J Mol Sci. 2021;23.
- 85. Dobson R, Giovannoni G. Multiple sclerosis - a review. Eur J Neurol. 2019;26:27–40.
- 86. Klineova S, Lublin FD. Clinical course of multiple sclerosis. Cold Spring Harb Perspect Med. 2018;8.
- 87. Montalban X, Gold R, Thompson AJ, Otero -Romero S, Amato MP, Chandraratna D, et al. ECTRIMS/EAN guideline on the pharmacological treatment of people with multiple sclerosis. Mult Scler. 2018;24:96–120.
- 88. Hauser SL, Cree BAC. Treatment of multiple sclerosis: a review. Am J Med. 2020;133(1380–90): e2.
- 89. Dangond F, Donnelly A, Hohlfeld R, Lubetzki C, Kohlhaas S, Leocani L, et al. Facing the urgency of therapies for progressive MS—a Progres ‑ sive MS Alliance proposal. Nat Rev Neurol. 2021;17:185–92.
- 90. Ferreira H, Amorim D, Lima AC, Pirraco RP, Costa -Pinto AR, Almeida R, et al. A biocompatible and injectable hydrogel to boost the efficacy of stem cells in neurodegenerative diseases treatment. Life Sci. 2021;287: 120108.
- 91. Valenza M, Birolini G, Cattaneo E. The translational potential of cholesterol -based therapies for neurological disease. Nat Rev Neurol. 2023;19:583–98.
- 92. Zuccato C, Valenza M, Cattaneo E. Molecular mechanisms and potential therapeutical targets in Huntington's disease. Physiol Rev. 2010;90:905–81.
- 93. Tosi G, Fano RA, Bondioli L, Badiali L, Benassi R, Rivasi F, et al. Investiga ‑ tion on mechanisms of glycopeptide nanoparticles for drug delivery across the blood -brain barrier. Nanomedicine (Lond). 2011;6:423–36.
- 94. Belletti D, Grabrucker AM, Pederzoli F, Menrath I, Vandelli MA, Tosi G, et al. Hybrid nanoparticles as a new technological approach to enhance the delivery of cholesterol into the brain. Int J Pharm. 2018;543:300–10.
- 95. Duan S, Yang J, Cui Z, Li J, Zheng H, Zhao T, et al. Seed amplifcation assay of nasal swab extracts for accurate and non -invasive molecular diagnosis of neurodegenerative diseases. Transl Neurodegener. 2023;12:13.