

Newly Recognized Spotted Fever Group *Rickettsia* as Cause of Severe Rocky Mountain Spotted Fever–Like Illness, Northern California, USA

Will S. Probert, Monica P. Haw, Aran C. Nichol, Carol A. Glaser, Sarah Y. Park, Laura E. Campbell, Kavita K. Trivedi, Hannah Romo, Megan E.M. Saunders, Anne M. Kjemtrup, Kerry A. Padgett, Jill K. Hacker

The incidence of spotted fever group (SFG) rickettsioses in the United States has tripled since 2010. Rocky Mountain spotted fever, the most severe SFG rickettsiosis, is caused by *Rickettsia rickettsii*. The lack of species-specific confirmatory testing obfuscates the relative contribution of *R. rickettsii* and other SFG *Rickettsia* to this increase. We report a newly recognized rickettsial pathogen, *Rickettsia* sp. CA6269, as the cause of severe Rocky Mountain spotted fever–like illness in 2 case-patients residing in northern California. Multilocus sequence typing supported the recognition of this pathogen as a novel *Rickettsia* genotype most closely related to *R. rickettsii*. Cross-reactivity observed for an established molecular diagnostic test indicated that *Rickettsia* sp. CA6269 might be misidentified as *R. rickettsii*. We developed a *Rickettsia* sp. CA6269–specific real-time PCR to help resolve this diagnostic challenge and better characterize the spectrum of clinical disease and ecologic epidemiology of this pathogen.

Rickettsioses are undifferentiated febrile illnesses, often accompanied by myalgia and rash, that are caused by intracellular gram-negative bacteria of the genus *Rickettsia*. Spotted fever group (SFG) *Rickettsia* are transmitted through the bite of ticks or mites and are grouped together on the basis of antigenic

and genetic similarities that distinguish this group from other *Rickettsia*, namely the typhus group. The incidence of SFG rickettsioses in the United States increased 3-fold from 2010 to 2018 (1). The SFG *Rickettsia* associated with human illness in the United States include the tickborne agents *R. rickettsii* (Rocky Mountain spotted fever [RMSF]), *R. parkeri* (*Rickettsia parkeri* rickettsiosis), and *Rickettsia* 364D (Pacific Coast tick fever [PCTF]) and the miteborne agent *R. akari* (rickettsialpox) (1,2). Among the SFG rickettsioses, RMSF is the most severe; its case-fatality rate is 5%–10% in the United States (3,4). Unfortunately, the relative contribution of *R. rickettsii* and other rickettsial agents to the increase in SFG rickettsioses is largely unknown because of the reliance on group-specific (SFG or typhus group) serologic testing and the lack of species-specific confirmatory testing (1,5).

In California, SFG rickettsioses are a reportable condition tracked for public health surveillance purposes. California has the lowest incidence of SFG rickettsioses among reporting states and averages only 11 probable and 1 confirmed SFG rickettsiosis cases annually (1,6,7). Only RMSF and PCTF are endemic to California (7). Ticks infected with *R. rickettsii* are extremely rare in California; only 1 infected *Dermacentor occidentalis* tick and 2 *Rhipicephalus sanguineus* ticks have been detected despite numerous environmental surveys (7). In contrast, *Rickettsia* 364D, a genetic near neighbor of *R. rickettsii*, is found in 2.1% of adult *D. occidentalis* ticks (8,9). PCTF was recognized in 2008 and is clinically less severe than RMSF (10). The presence of an inoculation eschar at the site of a tick bite and lack of a rash can be useful in differentiating PCTF from RMSF (9). Another genetic near neighbor of *R. rickettsii*, *Rickettsia* sp. CA6269, recently was reported

Author affiliations: California Department of Public Health, Richmond, California, USA (W.S. Probert, M.P. Haw, C.A. Glaser, H. Romo, M.E.M. Saunders, K.A. Padgett, J.K. Hacker); Kaiser Permanente, Oakland, California, USA (A.C. Nichol); Karius, Redwood City, California, USA (S.Y. Park); Alameda County Public Health Department, San Leandro, California, USA (L.E. Campbell, K.K. Trivedi); California Department of Public Health, Sacramento, California, USA (A.M. Kjemtrup)

DOI: <https://doi.org/10.3201/eid3007.231771>

in rabbit ticks (*Haemaphysalis leporispalustris*) collected in Northern California (11). Multilocus sequence typing (MLST) of that strain revealed a novel genotype having sufficient sequence divergence from *R. rickettsii* to lead the investigators to propose a new species, *Candidatus Rickettsia lanei*. Here, we describe the clinical and epidemiologic features of severe RMSF-like illness in 2 patients in northern California who had disease onset dates separated by nearly 20 years and whose illness was caused by a newly identified SFG *Rickettsia*, *Rickettsia* sp. CA6269.

Materials And Methods

Clinical Specimens

Clinical specimens for confirmatory testing of suspected rickettsial diseases were submitted to the California Department of Public Health Viral and Rickettsial Disease Laboratory (CDPH-VRDL) for public health surveillance purposes and considered exempt from human subject regulations by the California Health and Human Services Agency Committee for the Protection of Human Subjects (project no. 2023-085). In addition to a plasma specimen from the initial case-patient, archival clinical specimens (serum or plasma) collected over the past 20 years from 8 confirmed SFG rickettsiosis case-patients were available for molecular characterization. SFG rickettsiosis cases were classified as confirmed for public health surveillance purposes on the basis of clinical and laboratory criteria (12).

Nucleic Acid Extraction and Real-time PCR

We extracted total nucleic acids using the NucliSENS easyMAG instrument (bioMérieux, <https://www.biomerieux.com>) with a serum or plasma input volume of 300 μ L and a total nucleic acids output volume of 110 μ L. We further concentrated total nucleic acids from case-patient 2 by 4-fold using the RNA Clean and Concentrator Kit (Zymo Research, <https://www.zymoresearch.com>).

We tested specimens for *Rickettsia* using a laboratory-developed triplex real-time reverse transcription PCR (rRT-PCR) targeting a *R. rickettsii*-specific 23S rRNA single-nucleotide polymorphism (SNP), a *R. typhi*-specific 23S rRNA SNP, and genus-specific regions of the 23S rRNA (Appendix, <https://wwwnc.cdc.gov/EID/article/30/7/23-1771-App1.pdf>). The assay was developed and the performance specifications established by the CDPH-VRDL as required under the Clinical Laboratory Improvement Amendments. A *R. rickettsii*-specific real-time PCR (rPCR) assay, RRI6, was performed as described by Kato et al. (13); the assay does not detect *Rickettsia* 364D (14).

MLST

We amplified and sequenced 6 target sequences, 23S rRNA, 16S rRNA, *gltA*, *ompA*, *ompB*, and *sca4* (Appendix); the last 5 of those targets correspond to the regions recommended for the gene sequence-based classification of *Rickettsia* (15). A commercial laboratory performed bidirectional Sanger sequencing (ELIM Biopharmaceuticals, <https://elimbio.com>). We performed sequence editing and assembly using Geneious Prime 2022.0.2 (<https://www.geneious.com>). Nucleotide BLAST searches of the National Center for Biotechnology Information (NCBI) nucleotide collection and whole-genome shotgun contigs databases (<https://blast.ncbi.nlm.nih.gov>) were performed to identify genetic near neighbors and facilitate phylogenetic analyses. We concatenated and aligned the 16S rRNA, *gltA*, *ompA*, *ompB*, and *sca4* sequences for validly named SFG *Rickettsia* species and constructed a maximum-likelihood phylogenetic tree using MEGA 10 (<https://www.megasoftware.net>). We measured the reliability of the resulting tree by bootstrap resampling using 1,000 replicates.

Tick Collection and Testing

We conducted field investigations at potential exposure sites by flagging vegetation and leaf litter for ticks. We identified tick species, sex, and life stages by examining morphologic features. For the 2023 case investigation, we processed adult ticks collected from Alameda and Contra Costa Counties for nucleic acid extraction using the DNeasy Blood and Tissue Kit (QIAGEN, <https://www.qiagen.com>). We screened tick nucleic acid extracts for *R. rickettsii* using the RRI6 assay. Ticks collected and identified from San Mateo and Marin Counties during the 2004 case investigation were screened for *R. rickettsii* DNA by the US Army Center for Health Promotion and Preventative Medicine–West.

Design and Development of a *Rickettsia* sp. CA6269 Real-Time PCR

We performed a nucleotide BLAST search using the *ompA* sequence from *Rickettsia* sp. CA6269 (GenBank accession no. JN990595) and downloaded and aligned selected sequences using the multiple sequence alignment tool. We selected regions of sequence divergence for primer and probe design using Primer-BLAST (<https://www.ncbi.nlm.nih.gov/tools/primer-blast>). We designed a 5' exonuclease rPCR to amplify and detect a 146 bp region of *ompA*. The assay oligonucleotides were synthesized by Integrated DNA Technologies (<https://www.idtdna.com>). The 25- μ L reaction mixture consisted

of RLompAF1 (5'-GGGCACTTGGTGTTCCTACA-3') and RLompAR1 (5'-AAATGCCCAATTGTTTTGAG GAC-3') primers at 500 nM, RLompAPI (6FAM-CTAATGGTG/ZEN/ATCCTACTGGCG-3IABkFQ) probe at 100 nM, 1X Premix ExTaq (Probe qPCR) mix (Takara Biosciences, <https://www.takarabio.com>), and 5 µL of total nucleic acids. We performed amplification and fluorescence detection with the ABI 7500 FAST DX Sequence Detection System (ThermoFisher Scientific, <https://www.thermofisher.com>) using the fast mode and the following amplification conditions: 1 minute at 95°C, 45 cycles of 95°C for 3 seconds, and 60°C for 30 seconds. We assessed assay analytical specificity by using a panel of 37 total nucleic acid extracts from members of the order Rickettsiales (Appendix) and evaluated analytical sensitivity by using 10-fold serial dilutions of a quantified 156-bp gBlock gene fragment (Integrated DNA Technologies) spiked into nucleic acids from pooled human blood and tested in replicates of five. We defined the assay limit of detection as the smallest number of DNA copies detected per reaction for all 5 replicates.

Results

Case-Patient 1

The CDPH was notified of a suspected RMSF case involving a man with symptom onset in July 2023. The case-patient sought care at the emergency department (ED) with a 3-day history of influenza-like symptoms; he experienced fever, headaches, myalgias, arthralgias, malaise, loss of appetite, nonbloody diarrhea, and left-sided abdominal pain. Vital signs included temperature of 103°F (39.4°C), pulse of 96 beats/min, blood pressure of 116/71 mm Hg, and respiratory rate of 16 breaths/min. Lower left quadrant and suprapubic abdominal tenderness were noted upon physical examination. Of note, a rash was not observed. An antimicrobial regimen of ceftriaxone, metronidazole, and oral vancomycin was initiated, and the case-patient was hospitalized. The case-patient was transferred to the intensive care unit (ICU) on day 3 of hospitalization because of worsening hypoxemia, acidosis, encephalopathy, and seizures. Doxycycline was added to the treatment regimen on day 3 of hospitalization after an infectious diseases consultation that included rickettsial diseases in the differential diagnosis because of the severity of illness and clinical manifestations (Table 1).

Serologic testing for SFG *Rickettsia* by a commercial laboratory failed to detect a significant IgG titer in an acute serum specimen collected 5 days after symptom onset (Table 1). All blood cultures were negative.

A laboratory diagnosis of rickettsial infection was initially made with the Karius Test, a microbial cell-free DNA (mcfDNA) metagenomic sequencing method (16). Sequencing of a plasma specimen collected 7 days after the onset of symptoms determined that *R. rickettsii* detected at 254,523 DNA molecules/microliter and *R. slovaca* detected at 97,653 DNA molecules/microliter were the best matches in the Karius Test genomic reference database. Serologic testing of a convalescent serum specimen, collected 105 days after symptom onset, demonstrated a significant IgG titer of >1:256 for SFG *Rickettsia*. The case-patient was discharged after being hospitalized for 22 days, including 11 days in the ICU, with a primary diagnosis of RMSF and secondary diagnoses of septic shock, acute kidney injury caused by acute tubular necrosis requiring intermittent hemodialysis, acute hypoxemic respiratory failure, encephalopathy, seizures, diarrhea, hyponatremia, abnormal liver enzymes, thrombocytopenia, supraventricular tachycardia, atrial fibrillation, and gangrene involving multiple digits on both hands.

The residual plasma specimen from mcfDNA sequencing was provided to the CDPH-VRDL for species-specific confirmatory testing. We tested total nucleic acids extracted from this sample with the triplex rRT-PCR that detects a *Rickettsia*-specific sequence, an *R. rickettsii*-specific SNP, and an *R. typhi*-specific SNP in the 23S rRNA. Unexpectedly, only the *Rickettsia* species target, and not the *R. rickettsii*-specific SNP, was detected (cycle threshold [Ct] value 24.9) (Table 2). Subsequent testing with the RRI6 assay detected DNA at a Ct value of 30.5 (Table 2). To address the discordant *R. rickettsii* test results for the triplex rRT-PCR and the RRI6 assay, we sequenced a 1,468-nt segment of the rickettsial 23S rRNA (GenBank accession no. OR600926). Comparative sequence analysis revealed a 2-nt difference from *Rickettsia* 364D and a 3- to 4-nt difference from *R. rickettsii*, including an alternate SNP allele for the rRT-PCR SNP target. We further assessed species relatedness using a MLST scheme established for the classification of *Rickettsia* (15). We amplified and sequenced a 1,408 nt segment of 16S rRNA, a 1,051 nt segment of *gltA*, a 587 nt segment of *ompA*, a 4,849 nt segment of *ompB*, and a 2,944 nt segment of *sca4* (GenBank accession nos. OR600927 and OR596747–50). Searches of the NCBI nucleotide databases revealed perfect sequence matches with *gltA* (290 nt) and *sca4* (673 nt), a single nucleotide insertion/deletion difference in *ompA* (471 nt), and a single SNP difference in *ompB* (758 nt) from *Rickettsia* sp. CA6269 (GenBank accession nos. JN990594–7) (11). MLST results indicated that the strain, designated CA23RL1, was nearly

Table 1. Clinical and laboratory features of *Rickettsia* sp. CA6269 infections in study of newly recognized SFG *Rickettsia* as cause of severe RMSF-like illness, northern California, USA*

Feature	Case-patient 1	Case-patient 2
Clinical		
Fever at hospital admission	Present, 103°F (39.4°C)	Present, 101.8°F (38.8°C)
Inoculation eschar	Absent	Absent
Rash	Absent	Present, maculopapular including palms
Myalgia	Present	Present
Headache	Present	Present
Nausea	Present	Present
Vomiting	Present	Present
Diarrhea	Present	Absent
Abdominal pain	Present	Absent
Acute kidney injury	Present	Absent
Photophobia	Present	Present
Altered mental status	Present	Present
Acute respiratory failure	Present	Present
Cutaneous necrosis and gangrene	Present	Absent
Coma	Present	Present
Days in intensive care unit	11	4
Days hospitalized	22	13
Laboratory†		
Serum sodium	Low (124 mEq/L)	Low (133 mEq/L)
Serum lactate dehydrogenase	High (421 U/L)	High (1,280 U/L)
Serum creatinine	High (1.44 mg/dL)	Unremarkable (1.0 mg/dL)
Serum bilirubin, total	High (2.3 mg/dL)	Unremarkable (0.6 mg/dL)
Aspartate transaminase	High (483 U/L)	High (268 U/L)
Alanine transaminase	High (323 U/L)	High (125 U/L)
CBC platelets	Low (45K/uL)	Low (80K/uL)
CBC leukocyte count	Unremarkable (4.5K/uL)	High (12.9K/uL)
CBC neutrophil percentage	Unremarkable (64%)	High (84%)
CSF glucose	NA	Unremarkable (69 mg/uL)
CSF protein	NA	High (147 mg/dL)
CSF leukocyte count	NA	High (9/uL)
SFG <i>Rickettsia</i> IgG titer (acute)	Unremarkable <1:128	Elevated (1:4,096)
SFG <i>Rickettsia</i> IgG titer (convalescent)	Elevated (>1:256)	Elevated (1:16,384)

*CBC, complete blood count; CSF, cerebral spinal fluid; NA, not applicable; RMSF, Rocky Mountain spotted fever; SFG, spotted fever group.

†Samples for the blood metabolic panel, CBC, and CSF analyses (case-patient 2 only) were collected on the day of hospital admission for case-patient 1 and 3 days after hospital admission for case-patient 2.

identical to *Rickettsia* sp. CA6269. In contrast, CA23RL1 displayed significant sequence divergence from *R. rickettsii* and *Rickettsia* 364D at each MLST target (Table 3). Phylogenetic analysis of concatenated sequences from validly named SFG *Rickettsia* demonstrated that CA23RL1 occupied a distinct branch and was most closely related to *R. rickettsii* (Figure).

The case-patient had not traveled outside of the San Francisco Bay area within 19 days of illness onset and

did not recall a tick bite. The case-patient had played golf at 5 courses located in Alameda and Contra Costa Counties within 14 days of symptom onset, including the day symptoms began. Of note, the case-patient recalled entering vegetation to retrieve golf balls at several courses. Because that activity might have been a potential source of exposure to ticks, as follow-up we collected ticks from the 5 golf courses within 16–22 days (late July and early August) of illness onset. In all,

Table 2. Comparison of real-time PCR results for case-patient 1 and retrospective SFG rickettsiosis case-patients in study of newly recognized SFG *Rickettsia* as cause of severe RMSF-like illness, northern California, USA*

SFG rickettsiosis case-patient	Specimen type	Cycle threshold value, by test type				
		Triplex <i>Rickettsia</i> spp.	Triplex <i>R. rickettsii</i> SNP	Triplex <i>R. typhi</i> SNP	RRi6 <i>R. rickettsii</i>	<i>Rickettsia</i> sp. CA6269 ompA
1	Plasma	24.9	ND	ND	30.5	29.6
2	Serum	34.3	ND	ND	34.3	34.4
3	Plasma	23.3	25.2	ND	25.4	ND
4	Serum	30.3	27	ND	34	ND
5	Serum	32.8	30.2	ND	35	ND
6	Serum	32.2	32.8	ND	33.6	ND
7	Plasma	29.5	29.2	ND	33.3	ND
8	Serum	20.8	19.2	ND	25.1	ND
9	Plasma	28.6	27.3	ND	30.2	ND

*ND, not detected; RMSF, Rocky Mountain spotted fever; SFG, spotted fever group; SNP, single-nucleotide polymorphism.

Table 3. *Rickettsia* sp. CA23RL1 sequence similarity with *Rickettsia rickettsii* and *Rickettsia* 364D in study of newly recognized SFG *Rickettsia* as cause of severe RMSF-like illness, northern California, USA *

<i>Rickettsia</i> sp. CA23RL1 target sequence	<i>R. rickettsii</i> sequence similarity†	<i>Rickettsia</i> 364 sequence similarity‡
16S rRNA	99.7%	99.6%
<i>gltA</i>	99.2%	99.6%
<i>ompA</i>	96.8%	97.3%
<i>ompB</i>	98.6%	98.8%
<i>sca4</i>	98.6%	98.6%

*RMSF, Rocky Mountain spotted fever; SFG, spotted fever group.

†*Rickettsia rickettsii* strain Sheila Smith, GenBank accession no. CP000848.1.

‡*Rickettsia* 364D, GenBank accession no. CP003308.1.

197 adult ticks were collected and identified as either *D. occidentalis* (n = 135) or *D. similis* (n = 62). None of the ticks were infected with *R. rickettsii* or *Rickettsia* sp. CA6269 as determined by the RRI6 assay.

Case-Patient 2

We identified case-patient 2 through retrospective testing of 8 confirmed SFG rickettsiosis cases with the triplex rRT-PCR and RRI6 assays (Table 2). We detected *Rickettsia* spp. (Ct value 34.3), but not *R. rickettsii* or *R. typhi*, using the triplex rRT-PCR. The RRI6 assay detected rickettsial DNA (Ct value 34.3). Amplification and sequencing of a 397-nt segment of 23S rRNA (GenBank

accession no. OR600925) by nested RT-PCR revealed an exact match to the 23S rRNA sequence determined for case-patient 1 and a 2- to 4-nt difference from *Rickettsia* 364D and *R. rickettsii*, supporting the identification of a second case of *Rickettsia* sp. CA6269 infection.

Case-patient 2, a male adult, was diagnosed with RMSF in 2004 on the basis of clinical features and a 4-fold increase in serologic titer to SFG *Rickettsia* in serum specimens collected 10 days apart (Table 1). Illness onset occurred in June; the case-patient sought care at the ED with a 5-day history of headaches, vomiting, photophobia, neck pain, and confusion. Upon initial examination, vital signs included a body temperature of

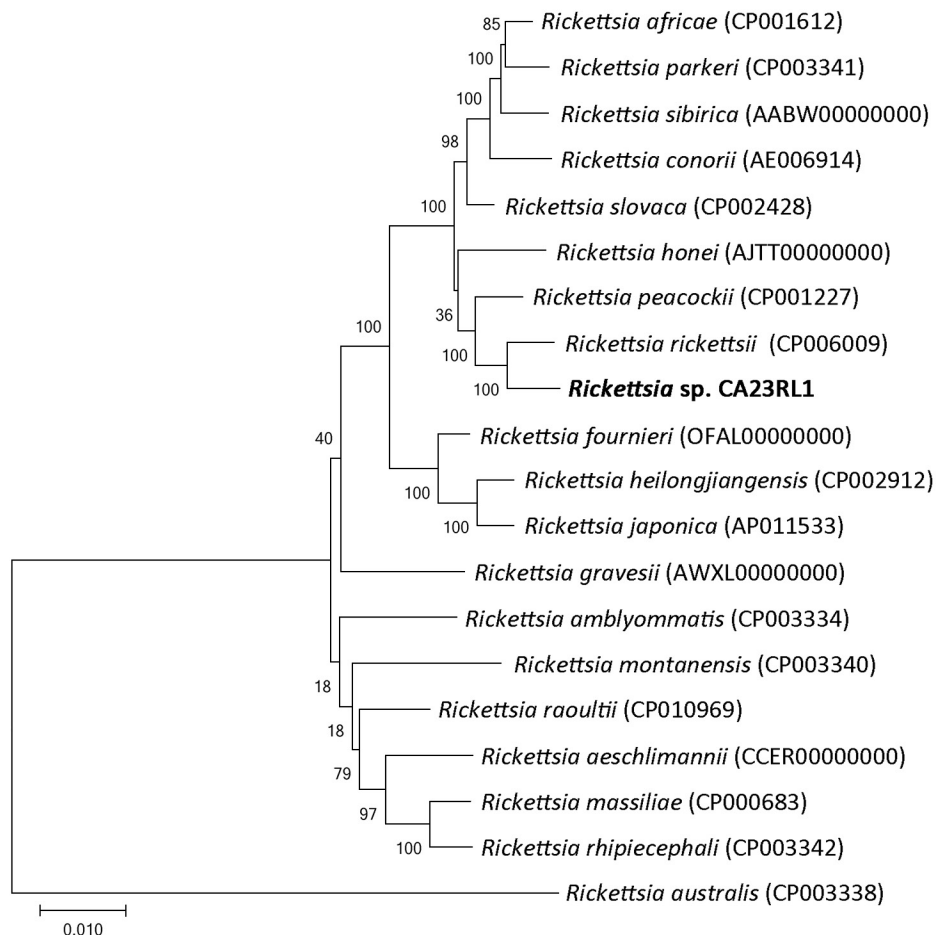


Figure. Maximum-likelihood phylogenetic tree of concatenated multilocus sequences in study of newly recognized spotted fever group *Rickettsia* as cause of severe Rocky Mountain spotted fever-like illness, northern California, USA. *Rickettsia* sp. CA23RL1 (bold text) occupies a distinct branch most closely related to *R. rickettsii*. *Rickettsia australis*, a transitional group *Rickettsia*, was included as an outgroup. Bootstrap values for 1,000 replicates are provided at each branch of the phylogenetic tree. GenBank accession numbers are provided in parentheses. Scale bar indicates evolutionary distance as measured by the number of substitutions per site.

101.8°F (38.8°C), pulse of 96 beats/min, blood pressure of 143/89 mm Hg, and respiratory rate of 20 breaths/min. Physical examination revealed mild nuchal rigidity, a maculopapular rash on the arms and legs, and leg edema. The case-patient was hospitalized on the basis of the ED assessment that included encephalitis, sepsis, hypoxemia, rash of unknown etiology, and chronic leg edema, and antimicrobial treatment with ceftriaxone, vancomycin, and acyclovir was initiated. On day 3 of hospitalization, the patient became hypoxic and comatose and was intubated and transferred to the ICU for 4 days. Doxycycline was added to the treatment regimen on day 3 of hospitalization after an infectious diseases consultation that included rickettsioses in the differential diagnosis on the basis of clinical manifestations and the patient's outdoor activities. Results of blood, CSF, urine, and sputum cultures were negative for significant pathogens. Serologic test results on day 10 of hospitalization indicated an IgG titer of 1:4,096 for SFG *Rickettsia*. After 13 days, the patient was discharged with a primary diagnosis of *Rickettsia* encephalitis.

The case-patient had not traveled outside the San Francisco Bay area during the 2 weeks before onset but had visited and camped at a county park and state beach in San Mateo County and a rural community in Marin County within that period. At the county park, he recalled finding a tick crawling on his body but not a tick bite. Field investigations at these locations conducted 30–40 days (July) after illness onset yielded 10 *D. occidentalis* ticks, 8 *D. similis* ticks, and 6 *Ixodes pacificus* ticks. Molecular testing of the *Dermacentor* spp. ticks did not detect *R. rickettsii*.

***Rickettsia* sp. CA6269–Specific Real-time PCR**

After the sequence-based identification of *Rickettsia* sp. CA6269 infections, we developed an rPCR targeting genotype-specific regions of *ompA*. We did not observe assay cross-reactivity with nucleic acids from *Anaplasma phagocytophilum*, *Ehrlichia chaffeensis*, *Orientia tsutsugamushi*, and 14 *Rickettsia* species, including 10 *R. rickettsii* strains and 11 *Rickettsia* 364D strains (Appendix). The assay limit of detection was 1 copy of DNA per reaction. Of 9 SFG rickettsiosis case-patients tested, *Rickettsia* sp. CA6269 was detected only for case-patient 1 (Ct value 29.6) and case-patient 2 (Ct value 34.4) (Table 2). The remaining 7 case-patients were confirmed *R. rickettsii* infections on the basis of detection of the species-specific 23S rRNA SNP with the triplex rRT-PCR.

Discussion

We describe the clinical and epidemiologic features of 2 case-patients with RMSF-like illness attributed to a newly recognized rickettsial pathogen, *Rickettsia* sp.

CA6269. The case-patients experienced severe clinical manifestations shared with RMSF, including acute kidney injury and respiratory failure, cutaneous necrosis and gangrene, and encephalitis. No unique clinical features were recognized in the 2 case-patients that would distinguish between infections caused by *Rickettsia* sp. CA6269 and *R. rickettsii*. Both patients likely acquired the infections locally during outdoor activities: in 1 case, golfing at courses with adjacent wildlands, and in the other case, visiting parks and camping. Although neither case-patient recalled being bitten by a tick, 1 case-patient had observed a tick crawling on his body. Nearly half of reported RMSF cases do not recall tick bites (4). These cases illustrate that clinicians should remain cognizant of SFG rickettsiosis in the differential diagnosis of undifferentiated febrile illnesses despite the lack of travel to areas of rickettsiosis endemicity and the absence of a recognized tick bite. Field investigations at locations of suspected exposure produced mostly *Dermacentor* spp. ticks and no *H. leporispalustris* ticks. Molecular testing of *Dermacentor* spp. ticks, established vectors of *R. rickettsii*, failed to detect *R. rickettsii* or *Rickettsia* sp. CA6269. Those potential locations of exposure will be the focus of future environmental investigations timed to better correlate with peak *H. leporispalustris* tick questing behavior.

Rickettsia sp. CA6269 was discovered during a survey of *H. leporispalustris* ticks collected in California for SFG *Rickettsia* (11). Of 234 ticks collected at the northern California site, 1 larval pool and 1 nymph produced a unique genotype, *Rickettsia* sp. CA6269. This unique genotype was not identified in the 179 ticks collected in southern California. The investigators proposed designating this novel *Rickettsia* as *Candidatus R. lanei* on the basis of results from an abbreviated MLST scheme. We have extended this work by sequencing the full-length regions originally proposed for the sequence-based classification of *Rickettsia* and established that *Rickettsia* sp. CA6269 meets the 2003 criteria for defining a new *Rickettsia* species (15). More recently, 2 genome sequence-based methods have been proposed for the classification of *Rickettsia* species (17,18). One method used genome sequence-based criteria aligned with the classic taxonomic analyses of bacteria to delineate *Rickettsia* into 9 species; nearly all SFG *Rickettsia* were classified as a single species (17). The other approach used genome sequence-based criteria that account for the unique phenotypic characteristics of rickettsiae and established phylogenetic relationships that were consistent with the current taxonomic classification of *Rickettsia* (18). That method displayed a strong correlation with MLST for *Rickettsia* classification. Despite the lack of updated consensus criteria for the taxonomic

classification of *Rickettsia* species, delineating *Rickettsia* sp. CA6269 as a strain or subspecies of *R. rickettsii* or a new species will benefit from isolating and cultivating strains, describing phenotypic characteristics including ecologic epidemiology, and determining whole-genome sequences.

Additional environmental studies are needed to investigate the geographic distribution, potential vectors, prevalence of infection, and reservoir hosts of *Rickettsia* sp. CA6269. The *H. leporispalustris* tick has been shown to be a maintenance vector of *R. rickettsii* and is distributed throughout the Americas from Alaska to Argentina (19,20). Adult *H. leporispalustris* ticks feed on rabbits and hares, whereas the nymph and larval stages feed on ground-frequenting birds and small rodents (21). Humans are rarely bitten by *H. leporispalustris*, indicating a limited role for this tick in transmitting SFG rickettsioses (22). Several surveys for *Rickettsia* infections in *H. leporispalustris* and rabbits have been conducted in California (11,23–26). In addition to the initial report of *Rickettsia* sp. CA6269, a further 2 studies have reported SFG *Rickettsia* detections in *H. leporispalustris*; 1 of those studies described the isolation of a *Rickettsia* antigenically related to *R. rickettsii* (11,23,24). Although likely rare, the case-patients described in this study might have encountered questing *H. leporispalustris* infected with *Rickettsia* sp. CA6269. Alternatively, *D. occidentalis*, a tick vector that more frequently bites humans and parasitizes a wide variety of mammals including lagomorphs, the preferred host of adult *H. leporispalustris*, might have been the source of *Rickettsia* sp. CA6269 transmission (27,28).

As with *R. rickettsii*, *Rickettsia* sp. CA6269 is likely a rarely encountered pathogen in California (7). After the initial detection of *Rickettsia* sp. CA6269, retrospective testing of 8 SFG rickettsiosis patient samples revealed a second case from nearly 20 years earlier. Nucleic acids from both cases lacked a *R. rickettsii*-specific 23S rRNA SNP but were detected with the RRI6 assay, indicating cross-reactivity of the RRI6 assay with *Rickettsia* sp. CA6269. The RRI6 assay was designed as *R. rickettsii*-specific on the basis of comparative genome analyses and targets a gene encoding a hypothetical protein absent in other rickettsial genomes including *Rickettsia* 364D (13,14). A homologue to this target sequence is likely present within the *Rickettsia* sp. CA6269 genome, creating the potential for misidentification of *Rickettsia* sp. CA6269 as *R. rickettsii* when using the RRI6 assay. To address this issue, we developed a *Rickettsia* sp. CA6269-specific rPCR assay that provides excellent analytical specificity and sensitivity. With more extensive performance characterization, this assay should prove useful for diagnostic testing and environmental

screening of potential vectors and reservoir hosts of *Rickettsia* sp. CA6269.

In summary, we have identified *Rickettsia* sp. CA6269 as a causative agent of severe RMSF-like illness in California and have described an assay for its detection. The application of this new test, both prospectively and retrospectively, could identify additional cases, thereby leading to a better understanding of both the clinical spectrum of disease caused by *Rickettsia* sp. CA6269 and the relative contribution of this pathogen to the increasing incidence of SFG rickettsioses in the United States. In addition, the new test will enable environmental studies to define the ecologic epidemiology of this emerging pathogen.

Acknowledgments

We thank Chris Paddock and Joy Hecht for providing the *Rickettsiaceae* nucleic acids and thank Summer Adams, Alexa Quintana, Ricardo Berumen, Sabrina Gilliam, Brandon Stavig, Bianca Gonzaga, and Maria Salas for technical and medical records support. We would also like to thank Darvin Scott Smith for participating in the California Encephalitis Project and facilitating specimen submission. We also gratefully acknowledge Mary Joyce Pakingan and Erin Trent for tick extraction, Contra Costa Mosquito and Vector Control District for tick collection, and Alameda County Vector Control Services District for both tick collection and extraction efforts.

This work is funded in part by the Centers for Disease Control and Prevention Epidemiology and Laboratory Capacity for Infectious Diseases program (grant Number 5 NU50CK000539).

The findings and conclusions in this article are those of the author(s) and do not necessarily represent the views or opinions of the California Department of Public Health or the California Health and Human Services Agency.

About the Author

Dr. Probert is a research scientist with the Viral and Rickettsial Disease Laboratory of the California Department of Public Health. He is interested in the development of molecular diagnostic assays for the detection and genotyping of microbial pathogens.

References

1. Bishop A, Borski J, Wang HH, Donaldson TG, Michalk A, Montgomery A, et al. Increasing incidence of spotted fever group rickettsioses in the United States, 2010–2018. *Vector Borne Zoonotic Dis.* 2022;22:491–7. <https://doi.org/10.1089/vbz.2022.0021>
2. Blanton LS. The rickettsioses: a practical update. *Infect Dis Clin North Am.* 2019;33:213–29. <https://doi.org/10.1016/j.idc.2018.10.010>

3. Paddock CD, Finley RW, Wright CS, Robinson HN, Schrodt BJ, Lane CC, et al. *Rickettsia parkeri* rickettsiosis and its clinical distinction from Rocky Mountain spotted fever. *Clin Infect Dis*. 2008;47:1188–96. <https://doi.org/10.1086/592254>
4. Biggs HM, Behravesh CB, Bradley KK, Dahlgren FS, Drexler NA, Dumler JS, et al. Diagnosis and management of tickborne rickettsial diseases: Rocky Mountain spotted fever and other spotted fever group rickettsioses, ehrlichioses, and anaplasmosis – United States. *MMWR Recomm Rep*. 2016;65:1–44. <https://doi.org/10.15585/mmwr.rr6502a1>
5. Binder AM, Nichols Heitman K, Drexler NA. Diagnostic methods used to classify confirmed and probable cases of spotted fever rickettsioses – United States, 2010–2015. *MMWR Morb Mortal Wkly Rep*. 2019;68:243–6. <https://doi.org/10.15585/mmwr.mm6810a3>
6. California Department of Public Health. IDB yearly summaries of selected communicable diseases in California, 2013–2021 [cited 2023 Oct 20]. <https://www.cdph.ca.gov/Programs/CID/DCDC/Pages/YearlySummSelectedGeneralCommDisinCA.aspx>
7. Kjemtrup AM, Padgett K, Paddock CD, Messenger S, Hacker JK, Feiszli T, et al. A forty-year review of Rocky Mountain spotted fever cases in California shows clinical and epidemiologic changes. *PLoS Negl Trop Dis*. 2022;16:e0010738. <https://doi.org/10.1371/journal.pntd.0010738>
8. Paddock CD, Yoshimizu MH, Zambrano ML, Lane RS, Ryan BM, Espinosa A, et al. *Rickettsia* species isolated from *Dermacentor occidentalis* (Acari: Ixodidae) from California. *J Med Entomol*. 2018;55:1555–60. <https://doi.org/10.1093/jme/tjy100>
9. Padgett KA, Bonilla D, Eremeeva ME, Glaser C, Lane RS, Porse CC, et al. The Eco-epidemiology of Pacific coast tick fever in California. *PLoS Negl Trop Dis*. 2016;10:e0005020. <https://doi.org/10.1371/journal.pntd.0005020>
10. Shapiro MR, Fritz CL, Tait K, Paddock CD, Nicholson WL, Abramowicz KF, et al. *Rickettsia* 364D: a newly recognized cause of eschar-associated illness in California. *Clin Infect Dis*. 2010;50:541–8. [10.1086/649926](https://doi.org/10.1086/649926) <https://doi.org/10.1086/649926>
11. Eremeeva ME, Weiner LM, Zambrano ML, Dasch GA, Hu R, Vilcins I, et al. Detection and characterization of a novel spotted fever group *Rickettsia* genotype in *Haemaphysalis leporispalustris* from California, USA. *Ticks Tick Borne Dis*. 2018;9:814–8. [10.1016/j.ttbdis.2018.02.023](https://doi.org/10.1016/j.ttbdis.2018.02.023) <https://doi.org/10.1016/j.ttbdis.2018.02.023>
12. Centers for Disease Control and Prevention. Spotted fever rickettsioses (including Rocky Mountain spotted fever) (SFR, including RMSF) 2020 case definition [cited 2023 Oct 20]. <https://ndc.services.cdc.gov/case-definitions/spotted-fever-rickettsiosis-2020>
13. Kato CY, Chung IH, Robinson LK, Austin AL, Dasch GA, Massung RF. Assessment of real-time PCR assay for detection of *Rickettsia* spp. and *Rickettsia rickettsii* in banked clinical samples. *J Clin Microbiol*. 2013;51:314–7. <https://doi.org/10.1128/JCM.01723-12>
14. Denison AM, Amin BD, Nicholson WL, Paddock CD. Detection of *Rickettsia rickettsii*, *Rickettsia parkeri*, and *Rickettsia akari* in skin biopsy specimens using a multiplex real-time polymerase chain reaction assay. *Clin Infect Dis*. 2014;59:635–42. <https://doi.org/10.1093/cid/ciu358>
15. Fournier PE, Dumler JS, Greub G, Zhang J, Wu Y, Raoult D. Gene sequence-based criteria for identification of new *rickettsia* isolates and description of *Rickettsia heilongjiangensis* sp. nov. *J Clin Microbiol*. 2003;41:5456–65. <https://doi.org/10.1128/JCM.41.12.5456-5465.2003>
16. Blauwkamp TA, Thair S, Rosen MJ, Blair L, Lindner MS, Vilfan ID, et al. Analytical and clinical validation of a microbial cell-free DNA sequencing test for infectious disease. *Nat Microbiol*. 2019;4:663–74. <https://doi.org/10.1038/s41564-018-0349-6>
17. Chung M, Munro JB, Tettelin H, Dunning Hotopp JC. Using core genome alignments to assign bacterial species. *mSystems*. 2018;3:e00236–18. <https://doi.org/10.1128/mSystems.00236-18>
18. Diop A, El Karkouri K, Raoult D, Fournier PE. Genome sequence-based criteria for demarcation and definition of species in the genus *Rickettsia*. *Int J Syst Evol Microbiol*. 2020;70:1738–50. <https://doi.org/10.1099/ijsem.0.003963>
19. Freitas LH, Faccini JL, Labruna MB. Experimental infection of the rabbit tick, *Haemaphysalis leporispalustris*, with the bacterium *Rickettsia rickettsii*, and comparative biology of infected and uninfected tick lineages. *Exp Appl Acarol*. 2009;47:321–45. <https://doi.org/10.1007/s100493-008-9220-4>
20. Kohls GM. Records and new synonymy of new world *Haemaphysalis* ticks, with descriptions of the nymph and larva of *H. juxtakochi* Cooley. *J Parasitol*. 1960;46:355–61. <https://doi.org/10.2307/3275499>
21. Burgdorfer W. Ecology of tick vectors of American spotted fever. *Bull World Health Organ*. 1969;40:375–81.
22. Nieto NC, Porter WT, Wachara JC, Lowrey TJ, Martin L, Motyka PJ, et al. Using citizen science to describe the prevalence and distribution of tick bite and exposure to tick-borne diseases in the United States. *PLoS One*. 2018;13:e0199644. [10.1371/journal.pone.0199644](https://doi.org/10.1371/journal.pone.0199644) <https://doi.org/10.1371/journal.pone.0199644>
23. Lane RS, Emmons RW, Dondero DV, Nelson BC. Ecology of tick-borne agents in California. I. Spotted fever group rickettsiae. *Am J Trop Med Hyg*. 1981;30:239–52. [10.4269/ajtmh.1981.30.239](https://doi.org/10.4269/ajtmh.1981.30.239) <https://doi.org/10.4269/ajtmh.1981.30.239>
24. Gurfieled N, Grewal S, Doggett D, Ferran K, LaFreniere R. Detection of spotted fever group *Rickettsia* and *Borrelia burgdorferi* in San Diego County rabbit ticks. *Proc Pap Annu Conf Mosq Vector Control Assoc Calif*. 2011;79:25–6.
25. Roth T, Lane RS, Foley J. A molecular survey for *Francisella tularensis* and *Rickettsia* spp. in *Haemaphysalis leporispalustris* (Acari: Ixodidae) in northern California. *J Med Entomol*. 2017;54:492–5. <https://doi.org/10.1093/jme/tjw202>
26. Schmitz KM, Foley JE, Kasten RW, Chomel BB, Larsen RS. Prevalence of vector-borne bacterial pathogens in riparian brush rabbits (*Sylvilagus bachmani riparius*) and their ticks. *J Wildl Dis*. 2014;50:369–73. <https://doi.org/10.7589/2012-11-292>
27. Eisen L. Tick species infesting humans in the United States. *Ticks Tick Borne Dis*. 2022;13:102025. <https://doi.org/10.1016/j.ttbdis.2022.102025>
28. Lane RS, Burgdorfer W. Spirochetes in mammals and ticks (Acari: Ixodidae) from a focus of Lyme borreliosis in California. *J Wildl Dis*. 1988;24:1–9. <https://doi.org/10.7589/0090-3558-24.1.1>

Address for correspondence: Will Probert, California Department of Public Health, Viral and Rickettsial Disease Laboratory, 850 Marina Bay Pkwy, Richmond, CA, 94804, USA; email: will.probert@cdph.ca.gov

Article DOI: <https://doi.org/10.3201/eid3007.231771>

EID cannot ensure accessibility for supplementary materials supplied by authors. Readers who have difficulty accessing supplementary content should contact the authors for assistance.

Newly Recognized Spotted Fever Group *Rickettsia* as Cause of Severe Rocky Mountain Spotted Fever–Like Illness, Northern California

Appendix

Triplex Real-time Reverse Transcription PCR (rRT-PCR)

Specimens were tested for *Rickettsia* using a laboratory-developed triplex real-time reverse transcription PCR (rRT-PCR) assay targeting a *R. rickettsii*-specific 23S rRNA single nucleotide polymorphism (SNP) (T101C, GenBank accession number NR_103963), a *R. typhi*-specific 23S rRNA SNP (T1976C, GenBank accession no. NR_076209), and genus-specific regions of the 23S rRNA (Table S1). The assay was developed and validated by the California Department of Public Health Viral and Rickettsial Disease Laboratory (validation report available upon request). The rRT-PCR mixture consisted of 1X One Step PrimeScript III RT-PCR master mix (Takara Bio USA, www.takarabio.com/), Rrick23S_F and Rrick23S_R primers at 300 nM, Rrick23S_P probe at 120 nM, Rtyp23S_F and Rtyp23S_R primers at 500 nM, Rtyp23S_P probe at 300 nM, and RCKr_F and RCKr_R primers at 500 nM, and RCKr_P at 80 nM. The nucleic acid input volume was 10 μ L for a final reaction volume of 25 μ L. Reverse transcription, amplification, and fluorescence detection were performed using an ABI 7500 FAST DX Sequence Detection System (Thermo Fisher Scientific, www.thermofisher.com/us/en/home.html) with the following cycling parameters: 50°C for 10 min, 95°C for 2 min followed by 40 cycles of 95°C for 3 s and 60°C for 40 s. Fluorescent readings were collected during the 60°C anneal/extension step.

References

- Chung IH, Robinson LK, Stewart-Juba JJ, Dasch GA, Kato CY. Analytically sensitive *Rickettsia* species detection for laboratory diagnosis. *Am J Trop Med Hyg.* 2022;106:1352–7. [PubMed](https://doi.org/10.4269/ajtmh.21-0757)
<https://doi.org/10.4269/ajtmh.21-0757>

Appendix Table 1. Triplex rRT-PCR oligonucleotide primer and probe sequences

Assay	Analyte	Oligonucleotide Name	Oligonucleotide Sequence and Modifications*
<i>Rickettsia</i> Triplex rRT-PCR	<i>Rickettsia</i> 23S rRNA†	RCKr_F	GGTCCYACAGACTTACCAAACCTCA
		RCKr_R RCKR_P	TCGACTATGGACCTTAGCACCCAT
		<i>R. rickettsii</i> 23S rRNA SNP	VIC-CCGAATGTCGATGAGTACAGCATAGCAGAC-QSY GCGATGAAGGACGTAATACGCT
<i>R. typhi</i> 23S rRNA SNP	SNP	Rrick23S_R	TAGGTAGGTTTCCCTATTCGGA
		Rrick23S_P	Q670-CGGATCGAAGTTTATTTCGCA-BHQ2
		Rtyp23S_F	CTAACGCCTCTGCTTCGCAG
		Rtyp23S_R	GAAAGACCCCGTGAACCTTTACTA
		Rtyp23S_P	6FAM-TGCACATTT/ZEN/GACTTCTAACACC-3IABkFQ

*Oligonucleotide modifications: 6FAM (6-carboxyfluorescein), VIC, Q670 (Quasar 670) are fluorescent dyes. QSY, BHQ2 (Black Hole Quencher 2), ZEN, and IABkFQ (Iowa Black Fluorescent Quencher) are nonfluorescent acceptor dyes.

†Modified from Chung et al. (1).

Appendix Table 2. Primer sequences, master mix, and amplification conditions for MLST

Amplification Reaction	Primer Name	Primer Sequences	Primer Reaction Concentration	Master Mix	Amplification Parameters
23S rRNA Outer RT-PCR Long Amplicon	RkAS77F2	CTTCGGGGAGTTGCGAATAA	600 nM	Superscript IV One-Step RT-PCR System	55°C for 20 min, 98°C for 2 min, 45 cycles of 98°C for 10 s, 64°C for 10 s, 72°C for 1 min, hold at 72°C for 5 min
	Rk23S1606R	GGTCATCTTTCTCCGAAGTTACAG	600 nM		
23S rRNA Inner PCR Long Amplicon	RkAS77F2	CTTCGGGGAGTTGCGAATAA	240 nM	Q5 Hot Start, High Fidelity	98°C for 30 s, 35 cycles of 98°C for 5 s, 64°C for 20 s, 72°C for 40 s, hold at 72°C for 2 min
	Rk23S1588R	GTTACAGATGYAATTTGCCTAGTT	240 nM		
23S rRNA Outer RT-PCR Short Amplicon	Rk23S42F	AGAAGGCGATGAAGGACGTAAT	600 nM	Superscript IV One-Step RT-PCR System	55°C for 20 min, 98°C for 2 min, 45 cycles of 98°C for 10 s, 62°C for 10 s, 72°C for 30 s, hold at 72°C for 5 min
	RkPan558R	GGTACGCCGTACAAGACAT	600 nM		
23S rRNA Inner PCR Short Amplicon	RkAS77F2	CTTCGGGGAGTTGCGAATAA	240 nM	Q5 Hot Start, High Fidelity	98°C for 30 s, 35 cycles of 98°C for 5 s, 64°C for 20 s, 72°C for 20 s, hold at 72°C for 2 min
	Rk23S517R	GCTTGTAAGCATTGGTTTCAGAT	240 nM		

Amplification Reaction	Primer Name	Primer Sequences	Primer Reaction Concentration	Master Mix	Amplification Parameters
16S rRNA Outer RT-PCR	Rk16S53F	AGTCTTTAAGGAGGTAATCCAGC	400 nM	Superscript IV One-Step RT-PCR System	55°C for 20 min, 98°C for 2 min, 45 cycles of 98°C for 10 s, 62°C for 10 s, 72°C for 1 min, hold at 72°C for 5 min
16S rRNA Inner PCR	Rk16S1570R	AACTGACAGAATCAAACCTTGAGAGT	400 nM	Q5 Hot Start, High Fidelity	98°C for 30 s, 35 cycles of 98°C for 5 s, 64°C for 30 s, 72°C for 30 s, hold at 72°C for 2 min
	Rk16S53F	AGTCTTTAAGGAGGTAATCCAGC	200 nM		
<i>gltA</i> PCR	Rk16S1556R SFGgltA71F	AACTTGAGAGTTTGATCCTGGCT GTTCCAGGCTTCGTGCATTC	200 nM 200 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 45 cycles of 98°C for 10 s, 64°C for 20 s, 72°C for 1 min, hold at 72°C for 2 min
<i>ompA</i> PCR	SFGgltA1226R RR190-70	AAAGCAAGTATCGGTGAGGATG ATGGCGAATATTTCTCCAAA	200 nM 200 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 45 cycles of 98°C for 10 s, 60°C for 20 s, 72°C for 40 s, hold at 72°C for 2 min
	RR190-701	GTTCCGTTAATGGCAGCATCT	200 nM		
<i>ompB1-2</i> Outer PCR	SFGompB65F	GAGCGATTAGAAGTTTACACGGA	200 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 45 cycles of 98°C for 10 s, 60°C for 20 s, 72°C for 1:30 min, hold at 72°C for 2 min
<i>ompB1</i> Inner PCR	SFGompB2850 R	CTACTACCGATGCTAACGTAGGT	200 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 35 cycles of 98°C for 5 s, 64°C for 20 s, 72°C for 40 s, hold at 72°C for 2 min
	SFGompB65F	GAGCGATTAGAAGTTTACACGGA			
<i>ompB2</i> Inner PCR	SFGompB1609 R	GCAAGTGGTACTTCAACATGGG	200 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 35 cycles of 98°C for 5 s, 64°C for 20 s, 72°C for 40 s, hold at 72°C for 2 min
	SFGompB1391 F	GTTAAATCTAGCACCACTTGGGA	200 nM		
<i>ompB3-4</i> Outer PCR	SFGompB2850 R	CTACTACCGATGCTAACGTAGGT	200 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 45 cycles of 98°C for 10 s, 60°C for 20 s, 72°C for 1:30
	SFGompB2692 F	CGTTAAATGTTGCATTACCTAAGAA CT	200 nM		

Amplification Reaction	Primer Name	Primer Sequences	Primer Reaction Concentration	Master Mix	Amplification Parameters
	SFGompB5000 R	AGGGTTGGTAACTGCTTCTACAG	200 nM		min, hold at 72°C for 2 min
<i>ompB3</i> Inner PCR	SFGompB2692 F	CGTTAAATGTTGCATTACCTAAGAA CT	400 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 35 cycles of 98°C for 5 s, 64°C for 20 s, 72°C for 40 s, hold at 72°C for 2 min
	SFGompB4180 R	GTTACAGGAAGTTTAGGCGGT	400 nM		
<i>ompB4</i> Inner PCR	SFGompB3546 F	CCGTTTATAACTGTACCGTCAGC	200 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 35 cycles of 98°C for 5 s, 64°C for 20 s, 72°C for 40 s, hold at 72°C for 2 min
	SFGompB5000 R	AGGGTTGGTAACTGCTTCTACAG	200 nM		
<i>sca4-1</i> PCR	SFGsca4-67F	TGAGAGGTTTTATGAGTAAAGACG GT	200 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 45 cycles of 98°C for 10 s, 64°C for 20 s, 72°C for 1 min, hold at 72°C for 2 min
	SFGsca4-1734R	CAATTGCTGCAGCTCTACTTGC	200 nM		
<i>sca4-2</i> PCR	SFGsca4-1533F	GCTGGAGTCAATGCAGTATTAGAA	200 nM	Q5 Hot Start, High Fidelity	98°C for 1 min, 45 cycles of 98°C for 10 s, 64°C for 20 s, 72°C for 1 min, hold at 72°C for 2 min
	SFGsca4-3158R	ACAGAATACAAATCTTGATCAGCGT	200 nM		

Appendix Table 3. MLST sequencing primers

Amplicon	Primer Name	Sequence
23S rRNA	RkAS77F2	CTTCGGGGAGTTGCCAATAA
	RkAS503F	CAAATGCTTACAAGCAGTCGGA
	Rk23S517R	GCTTGTAAGCATTGGTTTCAGAT
	Rk23S598R	GCTTGCTAGATAAACTAAGTCGC
	Rk23S1025F	GATGTGAGAAGACCAAAACAATA
	RkAS1158R	TCTTCGGTACATGACTTGAGCC
16S rRNA	Rk23S1588R	GTTACAGATGYAATTTGCC TAGTT
	Rk16S55F	AGTCTTTAAGGAGGTAATCCAGC
	Rk16S656F	TTATGCGTTAGCTGCGAAAC
	Rk16S709R	CGATGAGTGCTAGATATCGGAA
	Rk16S1556R	AACTTGAGAGTTTGATCCTGGCT
<i>gltA</i>	SFGgltA71F	GTTCAGGGTCTTCGTGCATTTTC
	RkgltA506F	GCATATGATGTTTGCAACGC
	RkgltA602R	CTCATGATCGGCATGTAGGA
<i>ompA</i>	SFGgltA1226R	AAAGCAAGTATCGGTGAGGATG
	RR190-70	ATGGCGAATATTTCTCCAAA
<i>ompB1</i>	RR190-701	GTTCCGTTAATGGCAGCATCT
	SFGompB67F	GAGCGATTAGAAGTTTACACGGA
<i>ompB2</i>	RkompB1-815F	GCTAGCGTATCTAAACCGATTAC
	RkompB1-893R	CACATCAAAGTAAGAAAGGTGGT
	SFGompB1609R	GCAAGTGGTACTTCAACATGGG
	SFGompB1391F	GTTAAATCTAGCACCACCTTGGG
	RkompB2-1996F	ACCGTCATTAATTGTTGCGTTA

Amplicon	Primer Name	Sequence
ompB3	RkompB2-2123R	ATACCCCTGGTACAGTTTATGG
	SFGompB2850R	CTACTACCGATGCTAACGTAGGT
	SFGompB2692F	CGTTAAATGTTGCATTACCTAAGAACT
	RkompB3-3479F	GCACTACCGTCTAAGGTAATAGT
ompB4	RkompB3-3592R	TGCCGGTTCTATCTTTAAACTTG
	SFGompB4180R	GTTACAGGAAGTTTAGGCGGT
	SFGompB3546F	CCGTTTATAACTGTACCGTCAGC
	RkompB4-4214F	AATACTAATCTACCCGTACCGTC
sca4-1	RkompB4-4278R	AATTTACAAGCAGGTGGTACT
	SFGompB5000R	AGGGTTGGTAACTGCTTCTACAG
	SFGsca4-67F	TGAGAGGTTTATGAGTAAAGACGGT
	RkSka4a758F	GCCGTATATTTCACTGCCCA
sca4-2	RkSka4a856R	TCCGGTCCGGCAAATTTTA
	SFGsca4-1734R	CAATTGCTGCAGCTCTACTTGC
	SFGsca4-1533F	GCTGGAGTCAATGCAGATTAGAA
	Rksca4-2322F	AGGTCCTGAATAACTAAGGCA
	Rksca4-2399R	TTAAGTGCTCTTTCTCAGAGT
	SFGsca4-3158R	ACAGAATACAAATCTTGATCAGCGT

Appendix Table 4. *Rickettsiales* nucleic acids used for assessing analytical specificity of the *Rickettsia* sp. CA6269 Real-time PCR Assay

Organism	Strain	Source
<i>Anaplasma phagocytophilum</i>	NCH-1	BEI Resources
<i>Ehrlichia chaffeensis</i>	St. Vincent	BEI Resources
<i>Orientia tsutsugamushi</i>	Karp	CDC
<i>Rickettsia africae</i>	R8637	CDC
<i>R. akari</i>	Kaplan	CDC
<i>R. amblyommatis</i>	GAT-30V	CDC
<i>R. asenbonensis</i>	NMRC11	CDC
<i>R. conorii</i>	R0362	CDC
<i>R. felis</i>	Baton Rouge	CDC
<i>R. massiliae</i>	AZT80	CDC
<i>R. parkeri</i>	Maculatum 20	CDC
<i>R. prowazekii</i>	Madrid II	CDC
<i>R. rhipicephali</i>	20-4164	CDC
<i>R. sibirica</i>	246	BEI Resources
<i>R. tillamookensis</i>	Tillamook 23	CDC
<i>R. typhi</i>	Wilmington	CDC
<i>R. rickettsii</i>	Hlp2	CDC
<i>R. rickettsii</i>	Columbia	CDC
<i>R. rickettsii</i>	Costa Rica	CDC
<i>R. rickettsii</i>	Brazil	CDC
<i>R. rickettsii</i>	Hauke	CDC
<i>R. rickettsii</i>	80JC	CDC
<i>R. rickettsii</i>	Coleman	CDC
<i>R. rickettsii</i>	Stewart	CDC
<i>R. rickettsii</i>	76RC	CDC
<i>R. rickettsii</i>	Sheila Smith	CDC
<i>Rickettsia</i> sp. 364D	Pierce Canyon	CDC
<i>Rickettsia</i> sp. 364D	Wright Peak	CDC
<i>Rickettsia</i> sp. 364D	Pine Mountain	CDC
<i>Rickettsia</i> sp. 364D	Crystal Cove	CDC
<i>Rickettsia</i> sp. 364D	El Moro Canyon	CDC
<i>Rickettsia</i> sp. 364D	Lake	CDC
<i>Rickettsia</i> sp. 364D	Cache Creek Ridge	CDC
<i>Rickettsia</i> sp. 364D	Highland Creek	CDC
<i>Rickettsia</i> sp. 364D	Orange	CDC
<i>Rickettsia</i> sp. 364D	Mt. Konocti	CDC
<i>Rickettsia</i> sp. 364D	364D	CDC