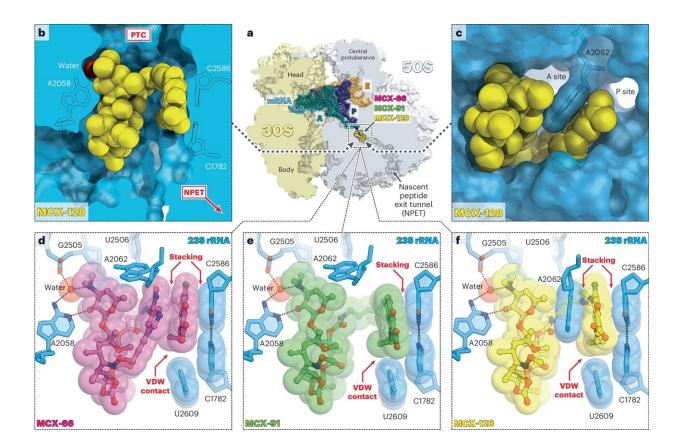


Dual action antibiotic could make bacterial resistance nearly impossible

July 23 2024, by Rob Mitchum



Structures of MCX-66, MCX-91 and MCX-128 in complex with the wild-type T. thermophilus 70S ribosome. Credit: *Nature Chemical Biology* (2024). DOI: 10.1038/s41589-024-01685-3

A new antibiotic that works by disrupting two different cellular targets would make it 100 million times more difficult for bacteria to evolve



resistance, according to new research from the University of Illinois Chicago.

For <u>a new paper</u> in *Nature Chemical Biology*, researchers probed how a class of synthetic drugs called macrolones disrupt bacterial cell function to fight infectious diseases. Their experiments demonstrate that macrolones can work two different ways—either by interfering with protein production or corrupting DNA structure.

Because <u>bacteria</u> would need to implement defenses to both attacks simultaneously, the researchers calculated that <u>drug resistance</u> is nearly impossible.

"The beauty of this antibiotic is that it kills through two different targets in bacteria," said Alexander Mankin, distinguished professor of pharmaceutical sciences at UIC. "If the antibiotic hits both targets at the same concentration, then the bacteria lose their ability to become resistant via acquisition of random mutations in any of the two targets."

Macrolones are synthetic antibiotics that combine the structures of two widely used antibiotics with different mechanisms. Macrolides, such as erythromycin, block the ribosome, the protein manufacturing factories of the cell. Fluoroquinolones, such as ciprofloxacin, target a bacteriaspecific enzyme called DNA gyrase.

Two UIC laboratories led by Yury Polikanov, associate professor of biological sciences, and Mankin and Nora Vázquez-Laslop, research professor of pharmacy, examined the cellular activity of different macrolone drugs.

Polikanov's group, which specializes in <u>structural biology</u>, studied how these drugs interact with the ribosome, finding that they bind more tightly than traditional macrolides. The macrolones were even capable of



binding and blocking ribosomes from macrolide-resistant bacterial strains and failed to trigger the activation of resistance genes.

Other experiments tested whether the macrolone drugs preferentially inhibited the ribosome or the DNA gyrase enzymes at various doses. While many designs were better at blocking one target or another, one that interfered with both at its lowest effective dose stood out as the most promising candidate.

"By basically hitting two targets at the same concentration, the advantage is that you make it almost impossible for the bacteria to easily come up with a simple genetic defense," Polikanov said.

The study also reflects the interdisciplinary collaboration at the UIC Molecular Biology Research Building, where researchers from the colleges of medicine, pharmacy and liberal arts and sciences share neighboring laboratories and drive basic science discoveries like this one, the authors said.

"The main outcome from all of this work is the understanding of how we need to go forward," Mankin said. "And the understanding that we're giving to chemists is that you need to optimize these macrolones to hit both targets."

In addition to Mankin, Polikanov and Vázquez-Laslop, UIC co-authors on the paper include Elena Aleksandrova, Dorota Klepacki and Faezeh Alizadeh.

More information: Elena V. Aleksandrova et al, Macrolones target bacterial ribosomes and DNA gyrase and can evade resistance mechanisms, *Nature Chemical Biology* (2024). DOI: <u>10.1038/s41589-024-01685-3</u>



Provided by University of Illinois at Chicago

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